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Role of lysophosphatidic acid receptor LPA₂ in the development of allergic airway inflammation in a murine model of asthma

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Abstract

Background: Lysophosphatidic acid (LPA) plays a critical role in airway inflammation through G protein-coupled LPA receptors (LPA₁₋₃). We have demonstrated that LPA induced cytokine and lipid mediator release in human bronchial epithelial cells. Here we provide evidence for the role of LPA and LPA receptors in Th2-dominant airway inflammation.

Methods:

Wild type, LPA₁ heterozygous knockout mice (LPA₁^{+/-}), and LPA₂ heterozygous knockout mice (LPA₂^{+/-}) were sensitized with inactivated *Schistosoma mansoni* eggs and local antigenic challenge with *Schistosoma mansoni* soluble egg Ag (SEA) in the lungs. Bronchoalveolar lavage (BAL) fluids and lung tissues were collected for analysis of inflammatory responses. Further, tracheal epithelial cells were isolated and challenged with LPA.

Results: BAL fluids from *Schistosoma mansoni* egg-sensitized and challenged wild type mice (4 days of challenge) showed increase of LPA level (~2.8 fold), compared to control mice. LPA₂^{+/-} mice, but not LPA₁^{+/-} mice, exposed to *Schistosoma mansoni* egg revealed significantly reduced cell numbers and eosinophils in BAL fluids, compared to challenged wild type mice. Both LPA₂^{+/-} and LPA₁^{+/-} mice showed decreases in bronchial goblet cells. LPA₂^{+/-} mice, but not LPA₁^{+/-} mice showed the decreases in prostaglandin E2 (PGE2) and LPA levels in BAL fluids after SEA challenge. The PGE2 production by LPA was reduced in isolated tracheal epithelial cells from LPA₂^{+/-} mice. These results suggest that LPA and LPA receptors are involved in *Schistosoma mansoni* egg-mediated inflammation and further studies are proposed to understand the role of LPA and LPA receptors in the inflammatory process.

Background

Lysophosphatidic acid (LPA) is a naturally occurring bioactive lysophospholipid and is a component of plasma, biological fluids, and tissues [1-3]. Many of the biological responses of LPA such as cell proliferation [4,5], migration [6,7], and cytokine release [8-10] are mediated by a family of G-protein coupled receptors (GPCRs). At least six LPA receptors, LPA₁₋₆, have been cloned and described in mammals, and the biological effects of LPA are mediated by ligation to specific LPA receptors that are coupled to heterotrimeric G-protein families, the G_s, G_i, G_q, and G_{12/13} [11-17].

The role of LPA and LPA receptors in airway inflammatory diseases have been studied *in vitro* and *in vivo*. LPA is a potent stimulator of interleukin-8 (IL-8) secretion in primary cultured human bronchial epithelial cells (HBEpCs) [8,10], and is a mitogen for airway smooth muscle cells [18,19]. Intratracheal administration of LPA in mice increased MIP-2 levels at 3 h and neutrophil infiltration at 6 h [20]. Inhalation of LPA induced histamine release and enhanced the recruitment eosinophils and neutrophils to the guinea pig lung alveolar space [21,22]. While these studies suggest that LPA regulates airway inflammation via stimulating the release of cytokines and inflammatory mediators that modulate infiltration of neutrophils and eosinophils into the airway, others point out that LPA exhibits anti-inflammatory effects and promotes resolution of inflammation. In human bronchial epithelial cells, LPA induced IL-13 decoy receptor, IL-13R α 2 expression and release, and attenuated IL-13-induced phosphorylation of STAT6 [9]. Further, LPA enhanced cyclooxygenase-2 (COX-2) expression and prostaglandin E2 (PGE2) release in HBEpCs [23] suggesting a protective role in the innate immunity response and tissue repair process in airway inflammation [24,25]. Recently, Fan et al. showed that intravenous injection with LPA attenuated bacterial endotoxin-induced plasma TNF- α production and myeloperoxidase activity in mouse lung, suggesting an anti-inflammatory role of LPA in a murine model of acute lung injury [26]. In addition to its anti-inflammatory effect, LPA regulated E-cadherin intracellular trafficking and airway epithelial barrier integrity and intratracheal post-treatment with LPA reduced neutrophil influx, protein leak, and E-cadherin shedding in bronchoalveolar lavage (BAL) fluids in a murine model of LPS-induced acute lung injury [27]. These data suggest a protective role of administered LPA in airway inflammatory diseases.

In contrast to several *in vitro* studies on the role of LPA as a pro- or anti-inflammatory mediator in airway epithelial and smooth muscle cells [8,10,18-20], there are a few reports linking LPA levels and LPA receptors to airway or lung inflammation and injury. We have recently shown that LPA was constitutively present in BAL fluids from normal and asthmatic subjects and segmental allergen

challenge increased LPA levels in BAL fluids significantly [28]. However, the source of LPA and the pathophysiological relevance of increased LPA after segmental allergen challenge to allergic inflammation remain to be elucidated. Similarly, LPA levels in BAL fluids from individuals with idiopathic pulmonary fibrosis were significantly higher compared to normal controls [29]. Further, an increase in LPA levels in BAL fluid following lung injury was observed in the bleomycin model of pulmonary fibrosis, and mice lacking LPA₁ were protected from fibrosis and mortality [29]. These studies suggest a role for LPA receptors in linking lung injury in the murine bleomycin model of pulmonary fibrosis.

Asthma is a chronic inflammatory disease of the airways involving T-lymphocytes and eosinophils infiltration, mucus overproduction and airway hyper-responsiveness. Inflammatory mediators including lipid mediators play a critical role in the pathogenesis of chronic airway diseases and facilitate the recruitment, activation, and trafficking of inflammatory cells in the airways. Very little is known on the physiological consequences of increased LPA levels and role of LPA receptors in asthma. To address the role of LPA receptors in Th2-mediated inflammation, we have used a well described *Schistosoma mansoni* eggs-sensitized murine model of allergic airway inflammation [30-32]. Control wild type, LPA₁^{+/-} and LPA₂^{+/-} mice were sensitized and challenged with *Schistosoma mansoni* eggs. LPA₂^{+/-} challenged mice compared to wild type showed decrease in cell numbers, eosinophils, and positive PAS staining. Interestingly, only *Schistosoma mansoni* eggs sensitized and challenged LPA₂^{+/-}, but not LPA₁^{+/-}, mice showed reduced PGE2 levels in BAL fluids which correlated with diminished COX-2 expression in LPA₂^{+/-} mice. Furthermore, airway epithelial cells isolated from LPA₂^{+/-} mice exhibited reduced COX-2 expression and PGE2 release compared to cells from wild type mice. These results show for the first time a role for LPA₂ in the development of airway inflammation and pathogenesis of asthma.

Materials and methods

Animals

All the mice were bred and housed in a specific pathogen-free barrier facility maintained by the University of Chicago Animal Resources Center. The studies reported here conform to the principles outlined by the Animal Welfare Act and the National Institutes of Health guidelines for the care and use of animals in biomedical research.

PCR genotyping of LPA₁^{+/-} and LPA₂^{+/-} mice

Extract-N-Amp Tissue PCR kit (Sigma Aldrich, S. Louis) was utilized for isolating genomic DNA from mouse tail and amplifying DNA fragments. The primers for LPA₁ and LPA₂ knockout mice were described as previous studies [33,34].

Schistosoma mansoni eggs sensitization and challenge

Schistosoma mansoni eggs sensitization and challenge to induce murine allergic airway disease were described before [31]. In brief, at day 0, mice (6-8 weeks) were immunized by i.p. injection of 5,000 inactivated *Schistosoma mansoni* eggs. At day 7, the mice were challenged with 10 µg of SEA by intratracheal aspiration. The mice were studied at day 11.

Analyses of BAL fluids

BAL fluids were performed by an intratracheal injection of 1 ml of PBS solution followed by gentle aspiration. The lavage was repeated twice to recover a total volume of 1.8-2.0 ml. The lavage was centrifuged and supernatant was processed for PGE2 or LPA measurement. The percentages of cell types in BAL fluids were determined by FACS analysis with cell type-specific markers.

Histology

Lungs were removed from mice and lobes were sectioned sagittally, embedded in paraffin, cut into 5-µm sections. Periodic Acid Schiff (PAS) staining were performed by Pathology Core Facility in The University of Chicago.

Antibodies and flow cytometry

Antibody to mouse CCR3 (clone 831101.111) was obtained from R&D Systems (Minneapolis, MN). Cells were fixed with 4% paraformaldehyde for 10 min and incubated with staining antibodies for 30 min at 4°C. The samples were washed and analyzed on a FACS LSR-II (Becton Dickinson).

Isolation of tracheal epithelial cells

Briefly, mice were euthanized and their tracheas were isolated and digested with 0.1% protease (Type XIV, Sigma) overnight at 4°C. The tracheal cell suspension were transferred to 15 ml tube and spun at 1500 rpm for 3 min at 4°C and were pooled in BEGM medium (Lonza, Walkersville, MD).

LPA measurement by mass spectrometry

Lipids in BAL were extracted as described before [28]. In brief, LPA levels were determined using liquid chromatography and tandem mass spectrometry (LC) with ABI-4000 Q-TRAP hybrid triple quadrupole/ion trap mass spectrometer (MS) coupled with an Agilent 1100 liquid chromatography system. Lipids were separated using methanol/water/HCOOH, 79/20/0.5, v/v, with 5 mM NH₄COOH as solvent A and methanol/acetonitrile/HCOOH, 59/40/0.5, v/v, with 5 mM NH₄COOH as Solvent B. LPA molecular species were analyzed in negative ionization mode with declustering potential and collision energy optimized for LPA.

PGE2 measurement

Mouse tracheal epithelial cells grown on 6-well plates were challenged with LPA for 3 h, medium were collected and centrifuged at 5,000 × g for 10 min at 4°C. The supernatant or BAL fluid supernatant were transferred to new 2.0 ml-eppendorf tubes and frozen in -80°C for later analysis. Measurement of PGE2 levels, as 13, 14-dihydro-15-keto PGE2, was carried out using a commercial ELISA kit according to manufacture's instruction.

RNA isolation and Real-time RT-PCR

Total RNA was isolated from cultured mouse tracheal epithelial cells using TRizol® reagent (Life Technology, Rockville, MD) according to the manufacturer's instructions. RNA was quantified spectrophotometrically and 1 µg of RNA was reversed transcribed using cDNA synthesis kit (Bio-Rad) and Real-time PCR and quantitative PCR were performed to assess expression of the COX-2, LPA₁, LPA₂, LPA₃, LPA₄, and LPA₅ using primers designed based on mouse mRNA sequences (Table 1.). Amplicon expression in each sample was normalized to its 18S RNA content. The relative abundance of target mRNA in each sample was calculated as 2 raised to the negative of its threshold cycle value times 10⁶ after being normalized to the abundance of its corresponding 18S, [e.g., 2^{-(Target Gene Threshold Cycle)}/2^{-(18S Threshold Cycle)} × 10⁶].

Western blotting

Equal amounts of protein (20 µg) were subjected to 10% SDS/PAGE gels, transferred to polyvinylidene difluoride membranes, blocked with 5% (w/v) BSA in TBST (25 mM Tris-HCl, pH 7.4, 137 mM NaCl and 0.1% Tween-20) for 1 h and incubated with anti-COX-2 antibody in 5% (w/v) BSA in TBST for 1-2 h at room temperature. The membranes were washed at least three times with TBST in 15 min intervals and then incubated with a rabbit horserad-

Table 1: Primers for mouse LPA receptors and COX-2

LPA ₁	Forward: 5'-TCAACCTGGTGACCTTTGTG-3' Reverse: 5'-GGTCCAGAACTATGCCGAGA-3'
LPA ₂	Forward: 5'-ATATTCCTGCCGAGATGCTG-3' Reverse: 5'-AAGCTGAGTAACGGGCAGAC-3'
LPA ₃	Forward: 5'-ATTGCCTCTGCAACATCTCG-3' Reverse: 5'-ATGAAGAAGGCCAGGAGGTT-3'
LPA ₄	Forward: 5'-ACTGCGTTCCTCACCAACAT-3' Reverse: 5'-CGATCGGAAGGGATAGACAA-3'
LPA ₅	Forward: 5'-GCTCCAGTGCCCTGACTATC-3' Reverse: 5'-CAGAGCGTTGAGAGGGAGAC-3'
COX-2	Forward: 5'-CCCCACAGTCAAAGACACT-3' Reverse: 5'-GGCACCAGACCAAAGACTTC-3'

ish peroxidase-conjugated secondary antibody (1: 3,000) for 1 h at room temperature. The membrane was developed with enhanced chemiluminescence detection system according to Manufacturer's instructions.

Statistical analysis

All results were subjected to statistical analysis using one-way ANOVA and, whenever appropriate, analyzed by Student-Newman-Keuls test. Data are expressed as means \pm S.D. of triplicate samples from at least three independent experiments and level of significance was taken to $P < 0.05$.

Results

Schistosoma mansoni eggs sensitization and challenge increases LPA levels in BAL fluids

To investigate the role of LPA receptors in pathogenesis of asthma, we quantified LPA levels in BAL fluids from control and SEA-challenged mice. Mice were sensitized by i.p. injection of 5,000 inactivated *Schistosoma mansoni* eggs. At day 7, mice were challenged with or without 10 μ g of SEA by intratracheal aspiration and at day 11, BAL fluids were collected (Fig. 1) and lipid were extracted and LPA levels in BAL fluids were measured by LC-MS/MS with C17:0 LPA as an internal standard. As shown in Table 2, LPA was detectable ($\sim 1254.3 \pm 357.0$ pmole/ml) in control mice (sensitized with inactivated *Schistosoma mansoni* eggs but not SEA challenged), and there was a ~ 2.8 fold increase in LPA levels ($\sim 3557.9 \pm 109.3$ pmole/ml) in *Schistosoma mansoni* eggs sensitized and challenged mice, compared to control mice. Unsaturated molecular species of LPA (18:1, 20:4, 22:5, and 22:6) were detected in BAL fluids of control mice, which increased significantly after SEA challenge. These results show for the first time, to our knowledge, increase in LPA during allergic lung inflammation in a murine model of asthma.

Schistosoma mansoni eggs sensitization and challenge-induced airway inflammation is dependent on LPA₁ and LPA₂

To determine the role of LPA receptors in airway inflammation mediated by *Schistosoma mansoni* eggs sensitiza-

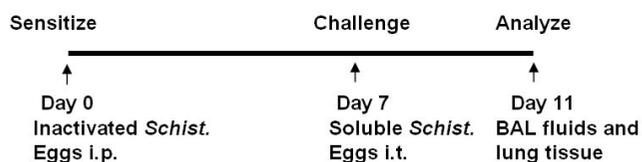


Figure 1
***Schistosoma mansoni* eggs sensitization and challenge induces murine asthmatic model.** At day 0, mice (6-8 weeks) were immunized by i.p. injection of 5,000 inactivated *Schistosoma mansoni* eggs. At day 7, mice were challenged with 10 μ g of SEA by intratracheal aspiration. Lung tissues and BAL fluids were collected at day 11.

tion and challenge, we used LPA₁ and LPA₂ deficient mice, which were genetically engineered as described earlier [33,34]. The heterozygous LPA₁^{+/-} and LPA₂^{+/-} mice were housed and bred at the University of Chicago Animal Resources Center and described experiments were approved by the ACIU of the University of Chicago. Genotyping analyses with specific primers confirmed generation of wild type (+/+), heterozygous (+/-) and homozygous mice (-/-) from the genetically engineered LPA₁ and LPA₂ mice (data not shown). Since LPA₁^{-/-} showed 50% neonatal lethality and impaired sucking in neonatal pups, all experiments were carried out with LPA₁^{+/-} and LPA₂^{+/-} mice to investigate the role of LPA receptors in *Schistosoma mansoni* eggs sensitization and challenge-mediated allergic inflammatory responses. To determine whether LPA₁^{+/-} and LPA₂^{+/-} mice reduced the effect of LPA, wild type, LPA₁^{+/-} and LPA₂^{+/-} mice were intratracheal challenged with 18:1LPA (5 μ M in 25 μ l PBS) for 6 h. As shown in Fig. 2, LPA challenge increased neutrophil infiltration, however, LPA₁^{+/-} and LPA₂^{+/-} mice reduced LPA-induced neutrophil infiltration in BAL fluids, suggesting that less LPA₁ and LPA₂ receptors in LPA₁^{+/-} and LPA₂^{+/-} mice reduce LPA-induced inflammation in lung and that LPA₁^{+/-} and LPA₂^{+/-} mice are useful models for investigating role of LPA receptors in lung inflammatory diseases.

Wild type, LPA₁^{+/-}, and LPA₂^{+/-} mice were sensitized with inactivated *Schistosoma mansoni* eggs and challenged with or without SEA for 4 days, BAL fluids and lung tissues were collected, cell numbers were measured under microscope and total eosinophils were determined by flow cytometry using eosinophils specific antibody (anti-CCR3). Consistent with previous reports [30,32], *Schistosoma mansoni* eggs sensitized and challenged wild type mice showed significant increase in total cell numbers and eosinophils in BAL fluids; however, total cell numbers and recruitment of eosinophils were attenuated in LPA₂^{+/-}, but not LPA₁^{+/-} mice (Fig. 3). These results suggest a role for LPA₂ in influx of eosinophils into alveolar space during allergic inflammatory response.

Airway goblet cell metaplasia and mucus production, indices of degree of inflammation, are hallmarks of asthma. Goblet cell metaplasia and mucus production were determined by PAS staining of histological sections of lung tissues from *Schistosoma mansoni* eggs sensitized and challenged or non-challenged wild type, LPA₁^{+/-}, and LPA₂^{+/-} mice. As shown in Fig. 4A, PAS positive goblet cells were higher in *Schistosoma mansoni* eggs sensitized and challenged wild type mice, compared to *Schistosoma mansoni* eggs sensitized and non-challenged wild type mice (control mice), whereas significantly less PAS stained goblet cells were seen in *Schistosoma mansoni* eggs sensitized and challenged LPA₁^{+/-} and LPA₂^{+/-} mice, compared to

Table 2: Quantification of LPA molecular species in BAL fluids

LPA molecular species	Wt sensitization only (pmol/ml)	Wt sensitization and SEA challenged (pmol/ml)
14:0-LPA	5.6 ± 3.1	14.6 ± 1.2
16:1-LPA	50.4 ± 37.0	160.9 ± 3.7
16:0-LPA	88.4 ± 51.7	253.6 ± 10.8
18:2-LPA	83.9 ± 54.6	274.4 ± 7.0
18:1-LPA	154.3 ± 114.7	517.6 ± 17.9
18:0-LPA	54.9 ± 27.4	153.7 ± 8.0
20:5-LPA	26.7 ± 22.0	95.7 ± 1.9
20:4-LPA	212.9 ± 168.2	739.0 ± 23.1
20:3-LPA	51.7 ± 39.8	165.2 ± 5.5
20:2-LPA	4.7 ± 3.4	14.1 ± 0.8
22:6-LPA	189.4 ± 114.4	548.9 ± 18.1
22:5-LPA	291.6 ± 166.2	576.9 ± 22.2
22:4-LPA	16.8 ± 9.5	41.1 ± 1.5
22:3-LPA	0.7 ± 0.6	1.9 ± 0.5
22:2-LPA	0.2 ± 0.1	0.2 ± 0.1
Total LPA	1254.3 ± 357.0	3557.9 ± 109.3

BAL fluids were collected and lipids were extracted. LPA molecular species were quantified by LC-MS/MS with 17:0LPA as standard.

Schistosoma mansoni eggs sensitized and challenged wild type mice. Scoring of the histological sections also confirmed a significantly higher percentage of bronchi for PAS positive stained cells in the sensitized and challenged control wild type mice compared to LPA₁^{+/-} and LPA₂^{+/-} mice (Fig. 4B). These results demonstrate that *Schistosoma mansoni* eggs sensitized and challenged LPA₁^{+/-} and LPA₂^{+/-} mice develop reduced goblet cell metaplasia and mucus production compared to control wild type mice. Together, these data suggest a role for LPA receptors for optimal induction of Th2-mediated airway inflammation.

LPA₂^{+/-}, but not LPA₁^{+/-}, mice exhibit reduced LPA and PGE2 levels in BAL fluids, and COX-2 expression in lungs of *Schistosoma mansoni* eggs sensitized/challenged mice

Endogenous PGE2 is produced by airway epithelium, smooth muscle, dendritic cells, and macrophages in response to allergen challenge [35]. PGE2 has been shown to be an anti-inflammatory lipid mediator and bronchodilator in the airway [24,25]; however, administration

of PGE2 induced various side effects, including cough, enhanced mucus production, and sensory nerve stimulation [36]. To determine the role of LPA receptors expression and PGE2 production in response to allergen challenge, we analyzed PGE2 levels in BAL fluids and COX-2 expression in lung tissues from *Schistosoma mansoni* eggs sensitized and challenged wild type mice. As shown in Fig. 5A, PGE2 levels were higher in control wild type and LPA₁^{+/-} mice, compared to LPA₂^{+/-} mice in response to *Schistosoma mansoni* eggs sensitization and challenge. *Schistosoma mansoni* eggs sensitization and challenge increased COX-2 expression in lung tissues of wild type mice while LPA₂^{+/-} mice showed reduced COX-2 expression (Fig. 5B). Recently, we have shown that LPA induces COX-2 expression and PGE2 release in human bronchial epithelial cells [23]. As *Schistosoma mansoni* eggs sensitization and challenge increased LPA levels in BAL fluids (Table 2), we measured LPA levels in BAL fluids from sensitized and SEA challenged LPA₂^{+/-} mice. Compared to wild type mice, LPA levels in BAL fluids from

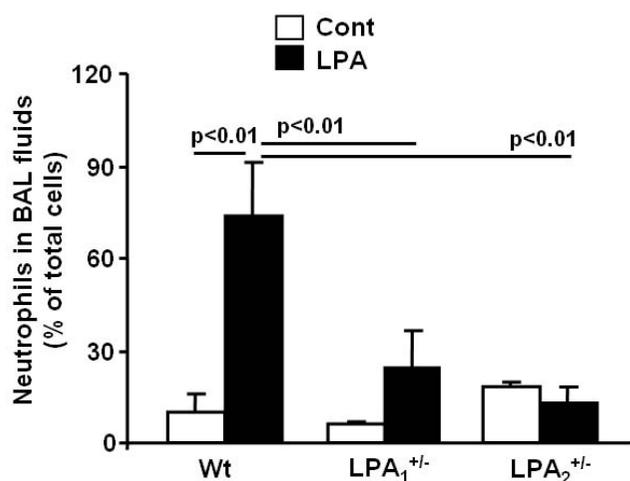


Figure 2
LPA₁^{+/-} and LPA₂^{+/-} mice show reduced neutrophils infiltration to BAL fluids. 18:1 LPA (5 μM in 25 μl PBS) were intratracheally injected to wild type, LPA₁^{+/-}, and LPA₂^{+/-} mice (n = 4-5) for 6 h. BAL fluids were collected and percentage of neutrophils in total cells were examined by Cytospin.

LPA₂^{+/-} mice were decreased after SEA challenge. Together, these results suggest that increased lung COX-2 expression, PGE2 and LPA production in BAL fluids by *Schistosoma mansoni* eggs sensitization and challenge is regulated by LPA₂.

LPA₂ deficiency on airway epithelial cells leads to reduced LPA mediated COX-2 expression and PGE2 release

Having demonstrated a role for LPA₂ in *Schistosoma mansoni* eggs-induced COX-2 expression, PGE2 secretion and airway inflammation, we hypothesized that expression of LPA₂ on airway epithelial cells may be involved in inflammatory responses to *Schistosoma mansoni* eggs sensitization and challenge. To investigate the role of LPA₂ in LPA-induced COX-2 expression and PGE2 production, tracheal epithelial cells were isolated from wild type and LPA₂^{+/-} mice. Analysis of total RNA for mRNA expression of LPA receptors by real-time RT-PCR revealed that expression of LPA₂>LPA₄>LPA₁≥LPA₃ in mouse tracheal epithelial cells (Table 3). In contrast to mouse tracheal epithelial cells, LPA₁ and LPA₃ were predominantly expressed in human bronchial epithelial cells [37]. In LPA₂^{+/-} tracheal epithelial cells, expression of LPA₂ mRNA was reduced to ~50%, compared to wild type mice, while there were no significant changes in expression levels of LPA₁ and LPA₃ mRNA (Fig. 6A). To determine the role of LPA₂ in LPA mediated COX-2 expression and PGE2 release, tracheal epithelial cells from wild type and LPA₂^{+/-} mice were challenged with LPA (1 μM) for 3 h, total RNA isolated and COX-2 mRNA expression determined by Real-time RT-

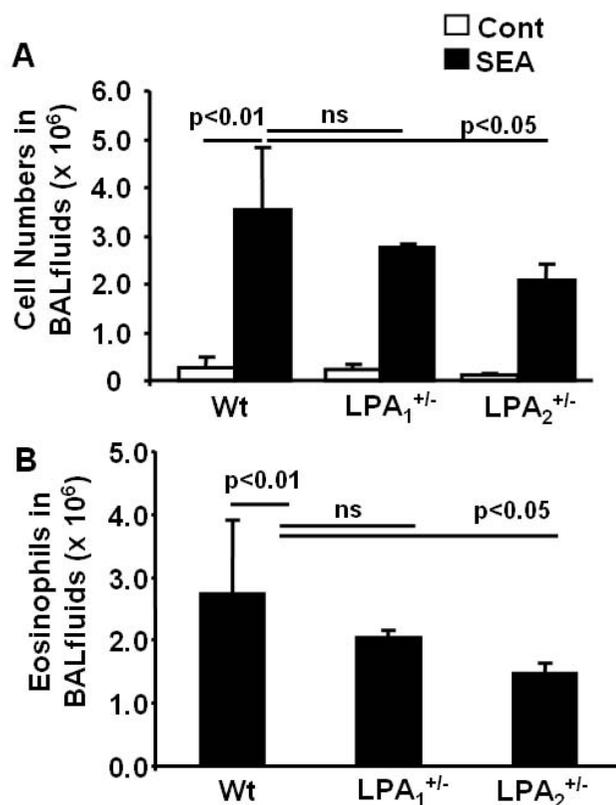


Figure 3
LPA₂^{+/-} mice exhibit a decrease in cell numbers and eosinophils in BAL fluids. After wild type, LPA₁^{+/-}, and LPA₂^{+/-} mice (n = 4-6) were challenged with or without *Schistosoma mansoni* eggs at day 11, as described in Materials and Methods, BAL fluids were collected and total cell numbers were accounted (A). Eosinophil numbers were examined by flow cytometry with antibody to CCR3 (B).

PCR. LPA stimulated COX-2 mRNA expression in wild type mouse cells (~13 fold); however, LPA-induced COX-2 mRNA expression was reduced in LPA₂^{+/-} mouse cells (~56% of wild type cells) (Fig. 6B). The media, after LPA challenge, were collected and PGE2 levels were determined. As shown in Fig. 6C, PGE2 release from LPA₂^{+/-} mouse tracheal epithelial cells challenged with LPA was lower as compared to cells from wild type mice [PGE2 (pg/ml)-Wild type: vehicle, 268 ± 29; LPA, 432 ± 47; LPA₂^{+/-}: vehicle, 283 ± 21; LPA, 374 ± 16]. These results suggest that a role for LPA₂ in LPA-induced COX-2 expression and PGE2 release from mouse tracheal epithelial cells.

Discussion

In the present study, we present several novel findings regarding LPA receptors expression, and its role in infiltration of eosinophils and lung inflammation in *Schistosoma mansoni* eggs sensitized and challenged murine model of

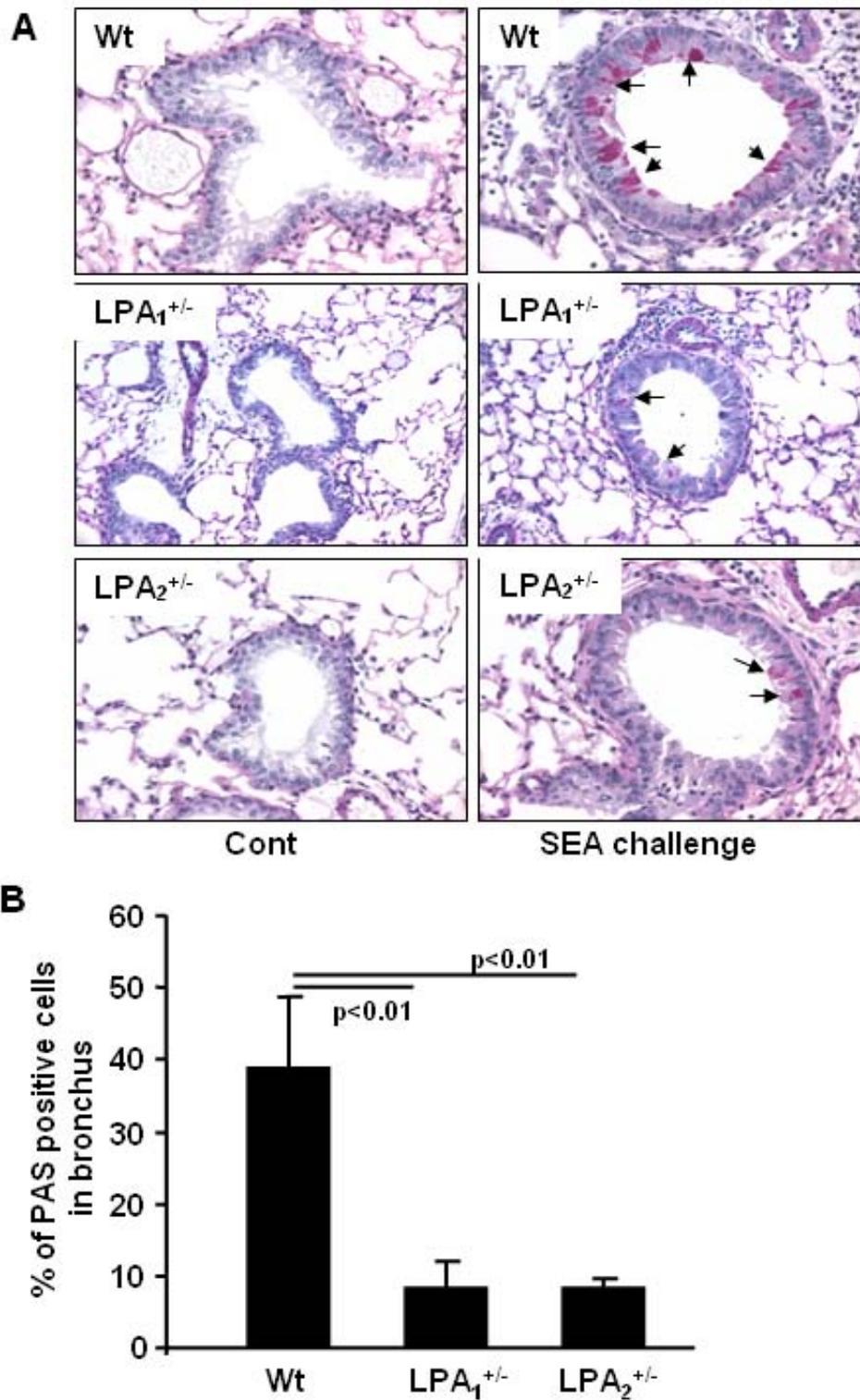


Figure 4
LPA₁^{+/-} and LPA₂^{+/-} mice exhibit decreases in goblet cells. **A)** Representative PAS staining sections from *Schistosoma mansoni* eggs unchallenged and challenged wild type, LPA₁^{+/-}, and LPA₂^{+/-} mice (n = 4-6) are shown. **B)** Percentage of PAS positive goblet cells in each bronchia (n = 3-5) were calculated.

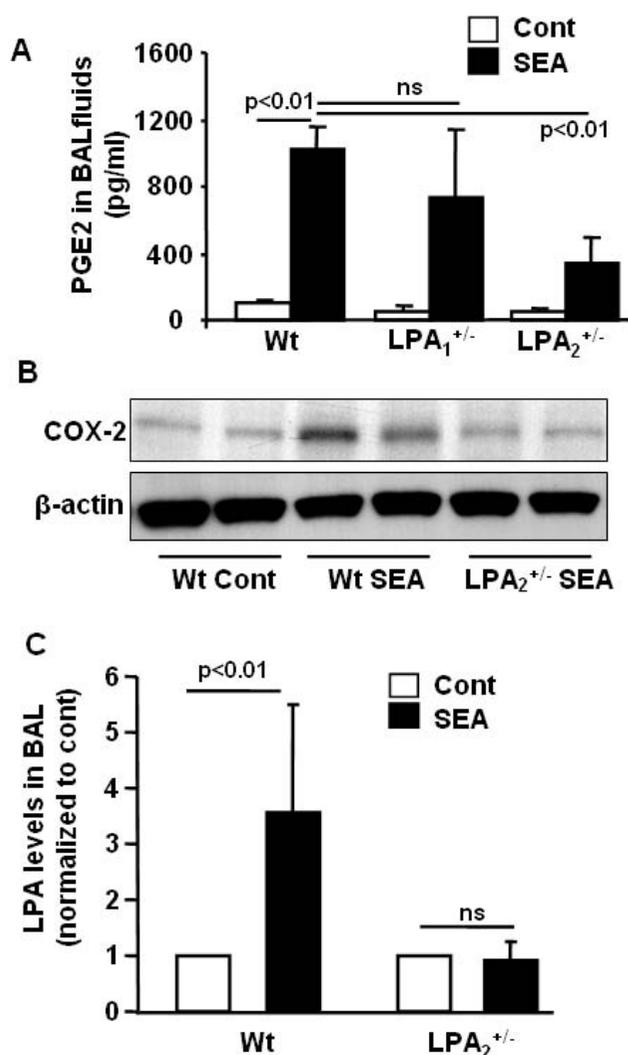
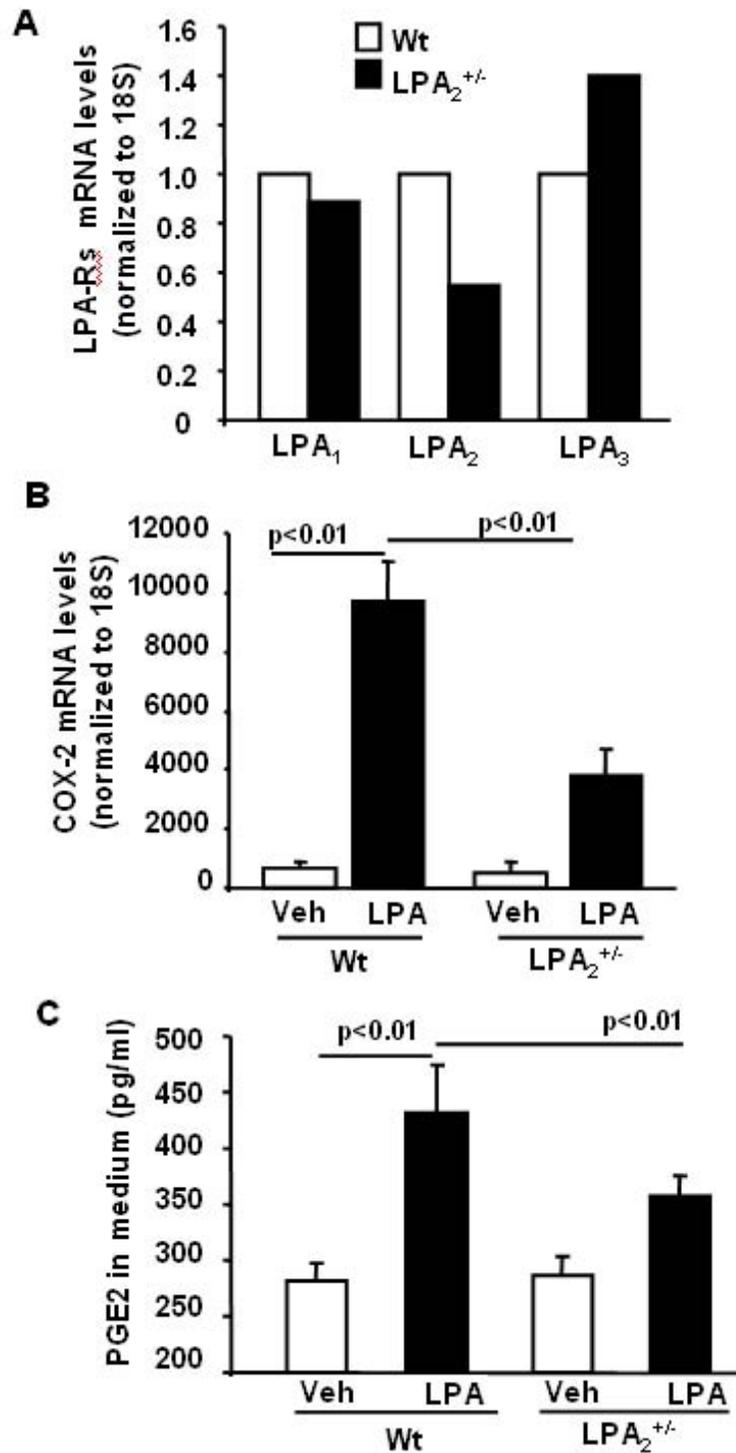


Figure 5
LPA₂^{+/-} mice exhibit a decrease in PGE2 and LPA levels in BAL fluids and COX-2 expression in lung tissue. BAL fluids and lung tissue were collected from SEA unchallenged and challenged wild type, LPA₁^{+/-}, and LPA₂^{+/-} mice (n = 4-6). **A)** PGE2 levels were measured by ELISA kit. **B)** Lung tissues were subjected to SDS/PAGE gel and COX-2 expression was determined by Western blotting. Representative image were shown. **C)** LPA levels in BAL fluids were quantified by LC-MS/MS and changes in LPA levels between wild type and SEA challenge mice were normalized to control levels.

asthma. We provide direct evidence for increased LPA levels in BAL fluids from *Schistosoma mansoni* eggs sensitized and challenged mice compared to control mice and a direct link between LPA₂ expression and lung inflammation mediated by *Schistosoma mansoni* eggs sensitization and challenge. The pro-inflammatory role of LPA₂ is also evident from reduced PGE2 levels in BAL fluids and COX-

2 expression in lung tissues of LPA₂^{+/-} mice sensitized and challenged with *Schistosoma mansoni* eggs compared to controls. We also demonstrate that airway epithelial cells isolated from LPA₂^{+/-} mice, compared to cells from wild type mice, exhibited reduced COX-2 expression and PGE2 release in response to LPA. To the best of our knowledge, this is the first report demonstrating a functional link between LPA, LPA₂ and lung inflammation in a murine model of asthma.

Asthma is a Th2-type immune disease of the lung that is characterized by chronic inflammation, infiltration of inflammatory cells, reversible obstruction of airway hyperresponsiveness, mucus hypersecretion by goblet cells and remodeling of the bronchoalveolar structures. Th1 and Th2 cytokines play a key role in orchestrating inflammatory and structural changes of the airway in asthma by recruiting, activating and promoting inflammatory cells into the airway [38-40]. In addition to cytokines, lipid mediators such as prostaglandins, leukotrienes, platelet-activating factor, and lysophospholipids regulate immune and inflammatory responses in asthma [41-43]. Many of these lipid mediators exert their biological responses via GPCRs. Increasing sphingosine-1-phosphate (S1P) levels in circulation offers protection against lung injury in mice and S1P-receptor 1 (S1P₁) heterozygous mice showed enhanced inflammation after LPS challenge suggesting an anti-inflammatory role of S1P₁ [44]. The present study demonstrates the role of LPA and LPA₂, a GPCR, in the pathogenesis of allergic airway inflammation in *Schistosoma mansoni* eggs sensitized and challenged murine model of asthma. LPA₁^{-/-} mice generated from LPA₁^{+/-} colonies, as compared to LPA₂^{-/-} from LPA₂^{+/-}, showed 50% neonatal lethality and impaired suckling, and therefore, we decided to use LPA₁^{+/-} and LPA₂^{+/-} mice to investigate role of LPA receptors in airway inflammation. Although LPA₁^{+/-} and LPA₂^{+/-} mice exhibited less neutrophils infiltration, compared to wild type mice, after LPA challenge (Fig. 2), influx of eosinophils was lower in LPA₂^{+/-}, but not in LPA₁^{+/-} mice after *Schistosoma mansoni* eggs sensitization and challenge (Fig. 3B). Both LPA₁^{+/-} and LPA₂^{+/-} mice showed reduced PAS positive cells in the bronchus compared to wild type after *Schistosoma mansoni* eggs sensitization and challenge (Fig. 4) suggesting the potential involvement of LPA₁ and LPA₂ in activation of goblet cells. These results indicate that activation of goblet cells are dependent on LPA₁ and LPA₂, however, only LPA₂ is involved in chemotaxis of eosinophils into alveolar space after *Schistosoma mansoni* eggs sensitization and challenge. Our current results on infiltration of eosinophils in *Schistosoma mansoni* eggs sensitized and challenged murine model of asthma are in good agreement with increased numbers of eosinophils, a characteristic feature of human bronchial asthma, in biopsies of human lung tissues [40,45]. LPA is constitutively

**Figure 6**

LPA induces COX-2 expression and PGE2 release through LPA₂. Tracheal epithelial cells from wild type and LPA₂^{+/-} mice were isolated as described in Materials and Methods and were cultured in 6-well plates. **A)** Total RNA was isolated and LPA receptors mRNA levels were measured by Real-time RT-PCR. **B)** Cells were challenged with 18:1LPA (1 μM) for 3 h, and COX-2 mRNA levels were measured by Real-time RT-PCR. **(C).** Cells were challenged with 18:1LPA (1 μM) for 3 h, and medium were collected. PGE2 levels in medium were measured by ELISA kit.

Table 3: LPA receptors mRNA expression in lung tissue

LPA-Rs	LPA ₁	LPA ₂	LPA ₃	LPA ₄	LPA ₅
Normalized to 18S	49.5 ± 23.5	235.0 ± 15.5	46.0 ± 21.2	73.5 ± 6.7	n.d.

Total RNA were isolated from wild type mouse tracheal epithelial cells and LPA receptors mRNA levels were determined by real-time RT-PCR with primers designed based on mouse LPA receptors mRNA sequence.

present in human BAL fluids and increased following allergic inflammation [28] and in patients with pulmonary fibrosis [29]. Intratracheal administration of LPA increased eosinophil influx in guinea pigs [22] and treatment of human eosinophils with LPA induced calcium mobilization, actin reorganization, and chemotaxis through G α i-dependent LPA receptors [46]. In the present study, we found that LPA levels were increased by ~3 fold following *Schistosoma mansoni* eggs sensitization and challenge of wild type, which supports the notion of LPA as a chemotaxis factor of inflammatory cells in allergic inflammation.

The source of LPA accumulation and mechanism(s) of LPA generation in the lung after allergic inflammation is unclear. Our previous studies have demonstrated that acyl glycerol kinase (AGK) converts monoacylglycerol to LPA in human bronchial epithelial cells [47]. Further, phospholipase D (PLD) can also contribute to intracellular LPA generation by providing phosphatidic acid, a substrate for PA specific phospholipase A2 [48,49]. Interestingly, we observed that LPA levels in LPA₂^{+/-} mice were significantly lower compared to wild type mice after *Schistosoma mansoni* eggs sensitization and challenge suggesting involvement of LPA₂ and potentially other LPA receptors in regulation of LPA generation in the airway. The relative contributions of AGK and/or PLD pathways in LPA generation in response to *Schistosoma mansoni* eggs sensitization and challenge are unknown. Additionally, extracellular LPA can be generated by lysoPLD (autotaxin), which converts lysophosphatidylcholine (LPC) to LPA [50]. Not only LPC levels were increased in BAL fluids of segmental allergen challenged patients [51], there was an increase in lysoPLD expression in LPS-stimulated monocytes [52], and stimulation of lysoPLD activity in asthmatic patients [53]. Thus, increase in LPC levels and lysoPLD expression and activity may be involved in enhanced LPA generation during lung inflammation. Further studies are needed to establish the potential source(s) of LPA in BAL fluids and mechanism(s) of LPA generation during allergic lung inflammation.

In contrast to LPA, there are only a few reports that describe the role of LPA receptors in lung inflammation, injury and remodeling. Deletion of LPA₁ reduced fibroblast recruitment and vascular leak in the bleomycin model of pulmonary fibrosis [29] while LPA/LPA₂ signaling via α v β 6 integrin-mediated activation of TGF- β has been

implicated in the development of bleomycin-induced lung fibrosis in mice [54]. Down-regulation of LPA₂ by siRNA attenuated LPA-induced phosphorylation of p38 MAPK/JNK, and IL-8 secretion in human bronchial epithelial cells [37]. Interestingly, *Schistosoma mansoni* eggs sensitization and challenge induced COX-2 expression and PGE2 was significantly attenuated in LPA₂^{+/-}, but not LPA₁^{+/-}, mice suggesting a potential link between reduced LPA₂ expression and COX2/PGE2 levels. In accordance with our *in vivo* results on *Schistosoma mansoni* eggs mediated COX-2 expression and PGE2 release in mouse lungs, tracheal epithelial cells from LPA₂^{+/-} mice exhibited decreased COX-2 expression and PGE2 release in response to LPA as compared to cells from wild type mice. Further, our results with LPA₂^{+/-} mice suggest a role for LPA₂ in the influx of eosinophils and lung inflammation induced by *Schistosoma mansoni* eggs sensitization and challenge suggest a role for LPA signaling via LPA₂ in pro-inflammatory responses.

Conclusion

The present study demonstrates increased LPA levels in BAL fluids in a murine model of asthma and LPA₂ heterozygous knockout mice show reduced Th-2 dominant airway inflammatory responses. These results suggest that endogenous LPA and LPA₂ play a critical role in pathogenesis of airway inflammatory diseases. Therapeutic targeting of LPA₂ may be beneficial in reducing allergic inflammatory responses in airway diseases.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

The study was designed and the protocol developed by YZ, JT, AIS, and VN. DH and SP carried out the genotyping. BE carried out the LPA measurement. JC provided the LPA₁ and LPA₂ heterozygous mice. All the authors read and approved the final manuscript.

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References

1. Aoki J, Taira A, Takanezawa Y, Kishi Y, Hama K, Kishimoto T, Mizuno K, Saku K, Taguchi R, Arai H: **Serum lysophosphatidic acid is**

- produced through diverse phospholipase pathways. *The Journal of biological chemistry* 2002, **277(50)**:48737-48744.
2. Postma FR, Jalink K, Hengeveld T, Bot AG, Alblas J, de Jonge HR, Moolenaar WH: **Serum-induced membrane depolarization in quiescent fibroblasts: activation of a chloride conductance through the G protein-coupled LPA receptor.** *The EMBO journal* 1996, **15(1)**:63-72.
 3. Tigyi G, Miledi R: **Lysophosphatidates bound to serum albumin activate membrane currents in Xenopus oocytes and neurite retraction in PC12 pheochromocytoma cells.** *The Journal of biological chemistry* 1992, **267(30)**:21360-21367.
 4. Mills GB, Moolenaar WH: **The emerging role of lysophosphatidic acid in cancer.** *Nature reviews* 2003, **3(8)**:582-591.
 5. van Corven EJ, Groenink A, Jalink K, Eichholtz T, Moolenaar WH: **Lysophosphatidate-induced cell proliferation: identification and dissection of signaling pathways mediated by G proteins.** *Cell* 1989, **59(1)**:45-54.
 6. Hao F, Tan M, Xu X, Han J, Miller DD, Tigyi G, Cui MZ: **Lysophosphatidic acid induces prostate cancer PC3 cell migration via activation of LPA(1), p42 and p38alpha.** *Biochimica et biophysica acta* 2007, **1771(7)**:883-892.
 7. van Leeuwen FN, Giepmans BN, van Meeteren LA, Moolenaar WH: **Lysophosphatidic acid: mitogen and motility factor.** *Biochemical Society transactions* 2003, **31(Pt 6)**:1209-1212.
 8. Zhao Y, He D, Saatian B, Watkins T, Spannake EW, Pyne NJ, Natarajan V: **Regulation of lysophosphatidic acid-induced epidermal growth factor receptor transactivation and interleukin-8 secretion in human bronchial epithelial cells by protein kinase Cdelta, Lyn kinase, and matrix metalloproteinases.** *The Journal of biological chemistry* 2006, **281(28)**:19501-19511.
 9. Zhao Y, He D, Zhao J, Wang L, Leff AR, Spannake EW, Georas S, Natarajan V: **Lysophosphatidic acid induces interleukin-13 (IL-13) receptor alpha2 expression and inhibits IL-13 signaling in primary human bronchial epithelial cells.** *The Journal of biological chemistry* 2007, **282(14)**:10172-10179.
 10. Zhao Y, Usatyuk PV, Cummings R, Saatian B, He D, Watkins T, Morris A, Spannake EW, Brindley DN, Natarajan V: **Lipid phosphate phosphatase-1 regulates lysophosphatidic acid-induced calcium release, NF-kappaB activation and interleukin-8 secretion in human bronchial epithelial cells.** *The Biochemical journal* 2005, **385(Pt 2)**:493-502.
 11. An S, Dickens MA, Bleu T, Hallmark OG, Goetzl EJ: **Molecular cloning of the human Edg2 protein and its identification as a functional cellular receptor for lysophosphatidic acid.** *Biochemical and biophysical research communications* 1997, **231(3)**:619-622.
 12. Chun J: **Lysophospholipid receptors: implications for neural signaling.** *Critical reviews in neurobiology* 1999, **13(2)**:151-168.
 13. Fukushima N, Kimura Y, Chun J: **A single receptor encoded by vzg-1/lpa1/edg-2 couples to G proteins and mediates multiple cellular responses to lysophosphatidic acid.** *Proceedings of the National Academy of Sciences of the United States of America* 1998, **95(11)**:6151-6156.
 14. Im DS, Heise CE, Harding MA, George SR, O'Dowd BF, Theodorescu D, Lynch KR: **Molecular cloning and characterization of a lysophosphatidic acid receptor, Edg-7, expressed in prostate.** *Molecular pharmacology* 2000, **57(4)**:753-759.
 15. Lee CV, Rivera R, Gardell S, Dubin AE, Chun J: **GPR92 as a new G12/13- and Gq-coupled lysophosphatidic acid receptor that increases cAMP, LPA5.** *The Journal of biological chemistry* 2006, **281(33)**:23589-23597.
 16. Noguchi K, Ishii S, Shimizu T: **Identification of p2y9/GPR23 as a novel G protein-coupled receptor for lysophosphatidic acid, structurally distant from the Edg family.** *The Journal of biological chemistry* 2003, **278(28)**:25600-25606.
 17. Tabata K, Baba K, Shiraiishi A, Ito M, Fujita N: **The orphan GPCR GPR87 was deorphanized and shown to be a lysophosphatidic acid receptor.** *Biochemical and biophysical research communications* 2007, **363(3)**:861-866.
 18. Cerutis DR, Nogami M, Anderson JL, Churchill JD, Romberger DJ, Rennard SI, Toews ML: **Lysophosphatidic acid and EGF stimulate mitogenesis in human airway smooth muscle cells.** *The American journal of physiology* 1997, **273(1 Pt 1)**:L10-15.
 19. Toews ML, Ustinova EE, Schultz HD: **Lysophosphatidic acid enhances contractility of isolated airway smooth muscle.** *J Appl Physiol* 1997, **83(4)**:1216-1222.
 20. Cummings R, Zhao Y, Jacoby D, Spannake EW, Ohba M, Garcia JG, Watkins T, He D, Saatian B, Natarajan V: **Protein kinase Cdelta mediates lysophosphatidic acid-induced NF-kappaB activation and interleukin-8 secretion in human bronchial epithelial cells.** *The Journal of biological chemistry* 2004, **279(39)**:41085-41094.
 21. Hashimoto T, Nakano Y, Yamashita M, Fang YI, Ohata H, Momose K: **Role of Rho-associated protein kinase and histamine in lysophosphatidic acid-induced airway hyperresponsiveness in guinea pigs.** *Japanese journal of pharmacology* 2002, **88(3)**:256-261.
 22. Hashimoto T, Yamashita M, Ohata H, Momose K: **Lysophosphatidic acid enhances in vivo infiltration and activation of guinea pig eosinophils and neutrophils via a Rho/Rho-associated protein kinase-mediated pathway.** *Journal of pharmacological sciences* 2003, **91(1)**:8-14.
 23. He D, Natarajan V, Stern R, Gorshkova IA, Solway J, Spannake EW, Zhao Y: **Lysophosphatidic acid-induced transactivation of epidermal growth factor receptor regulates cyclo-oxygenase-2 expression and prostaglandin E(2) release via C/EBPbeta in human bronchial epithelial cells.** *The Biochemical journal* 2008, **412(1)**:153-162.
 24. Oguma T, Asano K, Shiomi T, Fukunaga K, Suzuki Y, Nakamura M, Matsubara H, Sheldon HK, Haley KJ, Lilly CM, et al: **Cyclooxygenase-2 expression during allergic inflammation in guinea-pig lungs.** *American journal of respiratory and critical care medicine* 2002, **165(3)**:382-386.
 25. Vancheri C, Mastruzzo C, Sortino MA, Crimi N: **The lung as a privileged site for the beneficial actions of PGE2.** *Trends in immunology* 2004, **25(1)**:40-46.
 26. Fan H, Zingarelli B, Harris V, Tempel GE, Halushka PV, Cook JA: **Lysophosphatidic acid inhibits bacterial endotoxin-induced proinflammatory response: potential anti-inflammatory signaling pathways.** *Molecular medicine (Cambridge, Mass)* 2008, **14(7-8)**:422-428.
 27. He D, Su Y, Usatyuk PV, Spannake EW, Kogut P, Solway J, Natarajan V, Zhao Y: **Lysophosphatidic Acid Enhances Pulmonary Epithelial Barrier Integrity and Protects Endotoxin-induced Epithelial Barrier Disruption and Lung Injury.** *The Journal of biological chemistry* 2009, **284(36)**:24123-24132.
 28. Georas SN, Berdyshev E, Hubbard W, Gorshkova IA, Usatyuk PV, Saatian B, Myers AC, Williams MA, Xiao HQ, Liu M, et al: **Lysophosphatidic acid is detectable in human bronchoalveolar lavage fluids at baseline and increased after segmental allergen challenge.** *Clin Exp Allergy* 2007, **37(3)**:311-322.
 29. Tager AM, LaCamera P, Shea BS, Campanella GS, Selman M, Zhao Z, Polosukhin V, Wain J, Karimi-Shah BA, Kim ND, et al: **The lysophosphatidic acid receptor LPA1 links pulmonary fibrosis to lung injury by mediating fibroblast recruitment and vascular leak.** *Nature medicine* 2008, **14(1)**:45-54.
 30. Cannon JL, Collins A, Mody PD, Balachandran D, Henriksen KJ, Smith CE, Tong J, Clay BS, Miller SD, Sperling AI: **CD4 regulates Th2 differentiation and inflammation.** *J Immunol* 2008, **180(11)**:7385-7393.
 31. Padrid PA, Mathur M, Li X, Herrmann K, Qin Y, Cattamanchi A, Weinstock J, Elliott D, Sperling AI, Bluestone JA: **CTLA4lg inhibits airway eosinophilia and hyperresponsiveness by regulating the development of Th1/Th2 subsets in a murine model of asthma.** *American journal of respiratory cell and molecular biology* 1998, **18(4)**:453-462.
 32. Tong J, Bandulwala HS, Clay BS, Anders RA, Shilling RA, Balachandran DD, Chen B, Weinstock JV, Solway J, Hamann KJ, et al: **Fas-positive T cells regulate the resolution of airway inflammation in a murine model of asthma.** *The Journal of experimental medicine* 2006, **203(5)**:1173-1184.
 33. Contos JJ, Fukushima N, Weiner JA, Kaushal D, Chun J: **Requirement for the lpa1 lysophosphatidic acid receptor gene in normal suckling behavior.** *Proceedings of the National Academy of Sciences of the United States of America* 2000, **97(24)**:13384-13389.
 34. Contos JJ, Ishii I, Fukushima N, Kingsbury MA, Ye X, Kawamura S, Brown JH, Chun J: **Characterization of lpa(2) (Edg4) and lpa(1)/lpa(2) (Edg2/Edg4) lysophosphatidic acid receptor knockout mice: signaling deficits without obvious phenotypic abnormality attributable to lpa(2).** *Molecular and cellular biology* 2002, **22(19)**:6921-6929.

35. Newman JH, Sheller JR: **Arachidonic acid metabolites in the healthy and diseased lung.** *The American journal of the medical sciences* 1984, **288(2)**:75-80.
36. Gauvreau GM, Watson RM, O'Byrne PM: **Protective effects of inhaled PGE2 on allergen-induced airway responses and airway inflammation.** *American journal of respiratory and critical care medicine* 1999, **159(1)**:31-36.
37. Saatian B, Zhao Y, He D, Georas SN, Watkins T, Spannhake EW, Natarajan V: **Transcriptional regulation of lysophosphatidic acid-induced interleukin-8 expression and secretion by p38 MAPK and JNK in human bronchial epithelial cells.** *The Biochemical journal* 2006, **393(Pt 3)**:657-668.
38. Averbeck M, Gebhardt C, Emmrich F, Treudler R, Simon JC: **Immunologic principles of allergic disease.** *J Dtsch Dermatol Ges* 2007, **5(11)**:1015-1028.
39. Doherty T, Broide D: **Cytokines and growth factors in airway remodeling in asthma.** *Current opinion in immunology* 2007, **19(6)**:676-680.
40. Elsas PX, Elsas MI: **Eosinophilopoiesis at the cross-roads of research on development, immunity and drug discovery.** *Current medicinal chemistry* 2007, **14(18)**:1925-1939.
41. Bonnans C, Levy BD: **Lipid mediators as agonists for the resolution of acute lung inflammation and injury.** *American journal of respiratory cell and molecular biology* 2007, **36(2)**:201-205.
42. Peters-Golden M, Canetti C, Mancuso P, Coffey MJ: **Leukotrienes: underappreciated mediators of innate immune responses.** *J Immunol* 2005, **174(2)**:589-594.
43. Ryan JJ, Spiegel S: **The role of sphingosine-1-phosphate and its receptors in asthma.** *Drug news & perspectives* 2008, **21(2)**:89-96.
44. Sammi S, Vinasco LM, Mirzapioazova T, Singleton PA, Chiang ET, Evenoski CL, Wang T, Mathew B, Husain A, Moitra J, Sun X, Nenez L, Jacobson JR, Dudek S, Natarajan V, Garcia JGN: **Differential Effects of S1P Receptors on Airway and Vascular Barrier Function in the Murine Lung.** *American journal of respiratory and critical care medicine* 2009 in press.
45. Lee SY, Kim HB, Kim JH, Kim BS, Kang MJ, Jang SO, Hong SJ: **Eosinophils play a major role in the severity of exercise-induced bronchoconstriction in children with asthma.** *Pediatric pulmonology* 2006, **41(12)**:1161-1166.
46. Idzko M, Laut M, Panther E, Sorichter S, Durk T, Fluhr JW, Herouy Y, Mockenhaupt M, Myrtek D, Elsner P, et al.: **Lysophosphatidic acid induces chemotaxis, oxygen radical production, CD11b up-regulation, Ca2+ mobilization, and actin reorganization in human eosinophils via pertussis toxin-sensitive G proteins.** *J Immunol* 2004, **172(7)**:4480-4485.
47. Kalari S, Zhao Y, Berdyshev E, Usatyuk PV, He D, Natarajan V: **Role of acylglycerol kinase in lysophosphatidic acid-induced transactivation of epidermal growth factor receptor and expression of cyclooxygenase 2 and interleukin-8 in primary human bronchial epithelial cells.** *J Invest Medicine* 2007, **55(2)**:S356.
48. Aoki J: **Mechanisms of lysophosphatidic acid production.** *Seminars in cell & developmental biology* 2004, **15(5)**:477-489.
49. Sano T, Baker D, Virag T, Wada A, Yatomi Y, Kobayashi T, Igarashi Y, Tigyi G: **Multiple mechanisms linked to platelet activation result in lysophosphatidic acid and sphingosine 1-phosphate generation in blood.** *The Journal of biological chemistry* 2002, **277(24)**:21197-21206.
50. Morris AJ, Smyth SS: **Measurement of autotaxin/lysophospholipase D activity.** *Methods in enzymology* 2007, **434**:89-104.
51. Chilton FH, Averill FJ, Hubbard WC, Fonteh AN, Triggiani M, Liu MC: **Antigen-induced generation of lyso-phospholipids in human airways.** *The Journal of experimental medicine* 1996, **183(5)**:2235-2245.
52. Li S, Zhang J: **Lipopolysaccharide induces autotaxin expression in human monocytic THP-1 cells.** *Biochemical and biophysical research communications* 2009, **378(2)**:264-268.
53. Duff RF, Emo J, Friedman A, Larj M, Lyda E, O'Loughlin C, Rahman I, Block R, Morris A, Prestwich G, Georas SN: **Quantitative analysis of serum autotaxin levels in asthma and chronic obstructive pulmonary disease.** *American journal of respiratory and critical care medicine* 2009, **179**:A2528.
54. Xu MY, Porte J, Knox AJ, Weinreb PH, Maher TM, Violette SM, McAnulty RJ, Sheppard D, Jenkins G: **Lysophosphatidic acid induces alphavbeta6 integrin-mediated TGF-beta activation via the LPA2 receptor and the small G protein G alpha(q).** *The American journal of pathology* 2009, **174(4)**:1264-1279.

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