

**Enzymes** are catalysts evolved in nature to achieve the speed and coordination of a multitude of chemical reaction necessary to develop and maintain life.

**Enzymes** are globular proteins which range from 62 (monomer of 4-oxalocrotonate tautomerase) to over 2 500 amino acid residues (animal fatty acid synthase), but only a small portion (~ 3-4 amino acids are directly involved in catalysis)

### Classification

**Oxidoreductases** catalyze oxidation/reduction reactions

**Transferases** transfer a functional group (e.g. methyl group)

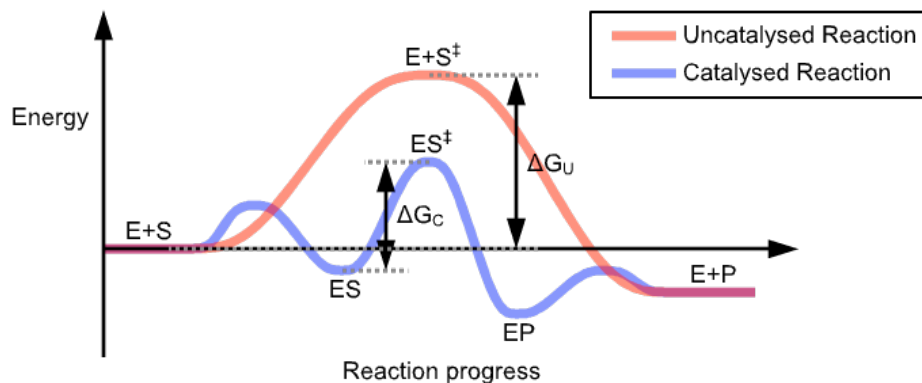
**Hydrolases** catalyze the hydrolysis of various bonds

**Lyases** cleave various bonds by means other than hydrolysis and oxidation

**Isomerases** catalyze isomerization changes within a single molecule

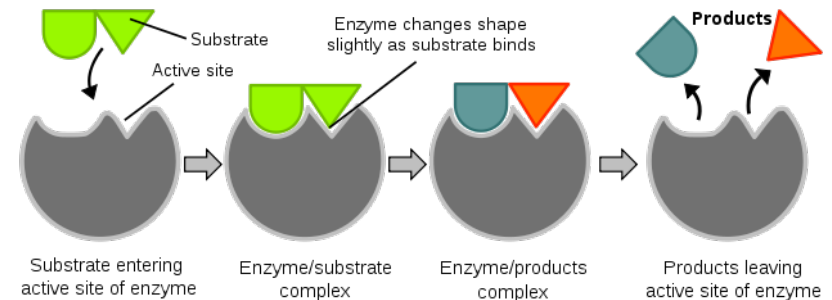
**Ligases** join two molecules with covalent bonds

### Enzyme reaction



### Induced fit

Model for the enzyme-substrate interaction introduced by Koshland



### Mechanism of transitions state stabilization

#### Catalysis by bond strain

affinity of the enzyme to the transition state is greater than to the substrate itself  $\rightarrow$  ground state destabilization effect

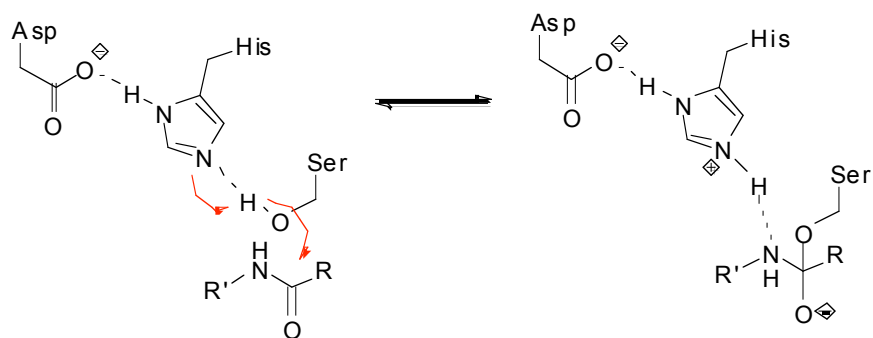
#### Catalysis by proximity and orientation

enzyme-substrate interactions align reactive groups and hold them close together  $\rightarrow$  reduces the overall loss of entropy

#### Catalysis involving proton donors or acceptors (acid/base catalysis)

stabilization of developing charges in the transition state  $\rightarrow$  activation of nucleophiles and electrophiles or stabilization of leaving groups

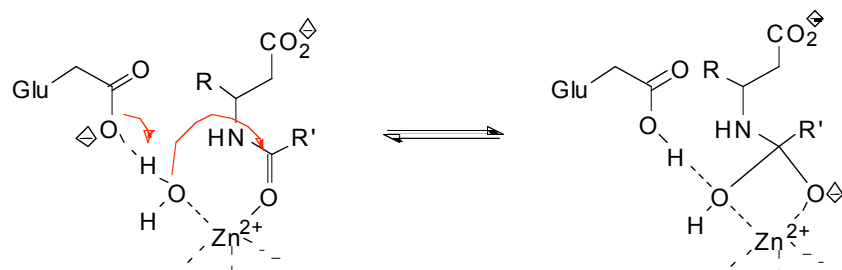
initial step of the serine protease catalytic mechanism:



### Electrostatic catalysis

stabilization of charged transition states by forming ionic bonds with residues of the active site

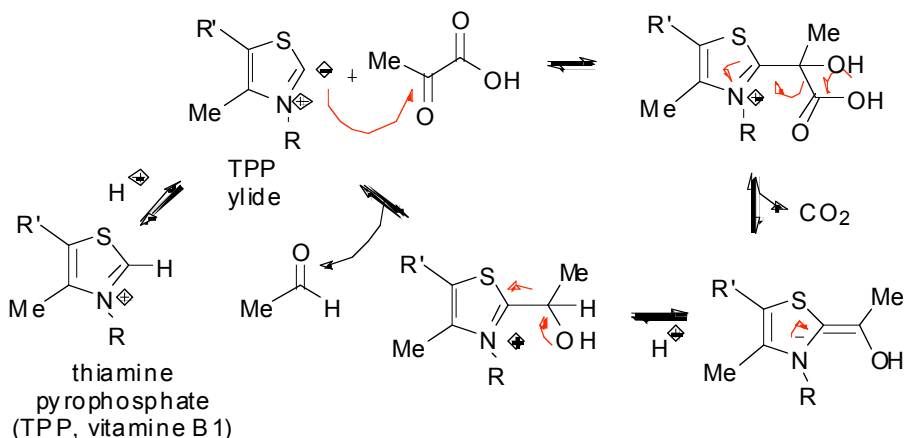
initial step of the carboxypeptidase catalytic mechanism:



### Covalent catalysis

substrate is forming a transient covalent bond with a residue in the active site in order to reduce energy of later transition states of the reaction

pyruvate decarboxylase mechanism



### Advantages of biocatalysts and enzymes

- very high enantioselectivity
- very high regioselectivity
- transformation under mild conditions
- 'green chemistry' e.g. solvent often water

### Disadvantages of biocatalysts and enzymes

- often low specific activity
- instability at extreme temperatures and pH values
- availability for selected reactions only
- long development time for new enzymes

advances in genomics, directed evolution, gene and genome shuffling and the exploration of Earth's biodiversity aided by bioinformatics and high-throughput screening facilitate the discovery and optimization of enzymes

**It is estimated that biocatalysis and biotransformations account for 30% of the chemical business by the year 2050**

## Biotransformations on an Industrial Scale

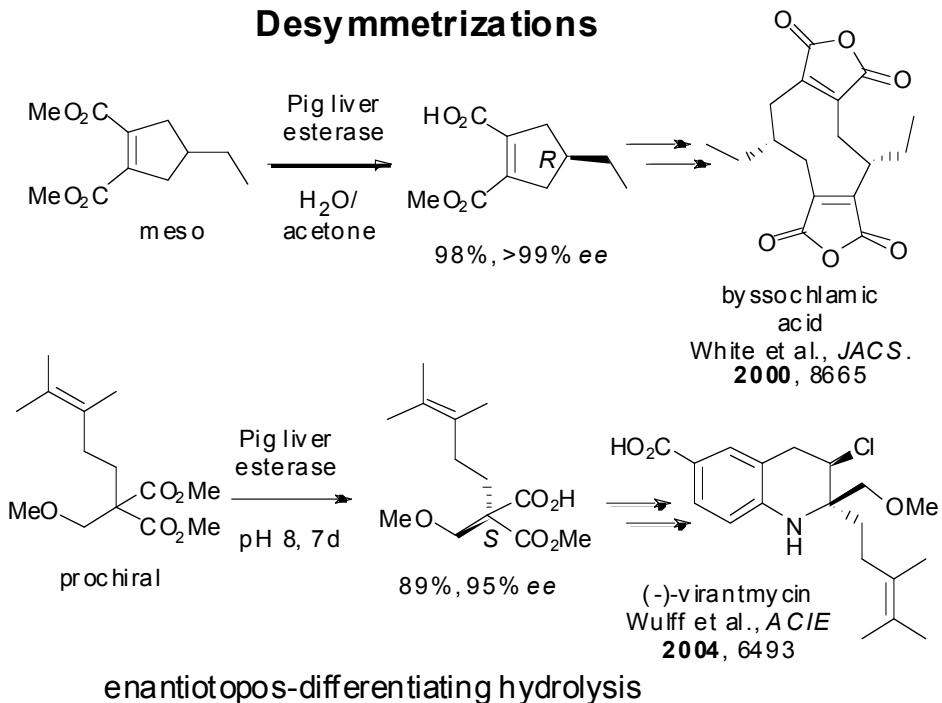
t/a	product	enzyme
> 1 000 000	high-fructose corn syrup	glucose isomerase
> 100 000	lactose-free milk	lactase
> 10 000	acrylamide	nitrilase
	cocoa butter	lipase
> 1 000	nicotinamide	nitrilase
	D-pantothenic acid	aldonolactonase
	(S)-chloropropionic acid	lipase
	6-aminopenillanic acid	penicillin amidase
	7-aminocephalosporanic acid	glutaryl amidase
	aspartame	thermolysin
	L-aspartate	aspartase
	D-phenylglycine	hydantoinase
	D- <i>p</i> -OH-phenylglycine	hydantoinase
> 100	ampicillin	penicillin amidase
	L-methionine, L-valine	aminoacylase
	L-carnitine	dehydrase/ hydroxylase
	L-DOPA	$\beta$ -tyrosinase
	L-malic acid	fumarase
	(S)-methoxyisopropyl-amine	lipase
	( <i>R</i> )-mandelic acid	nitrilase
	L-alanine	L-aspartate- $\beta$ -de-carboxylase

further applications:  
baby foods, brewing industry, fruit juice, dairy industry, starch,  
paper, biofuels, detergents, rubber,...

## Literature

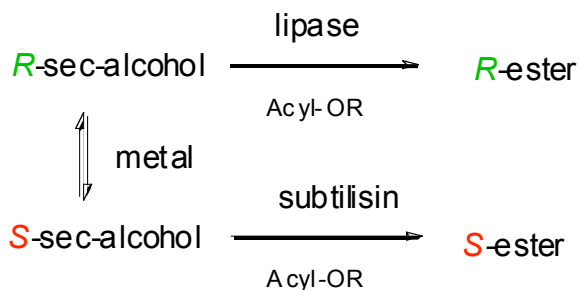
- K. Drauz, H. Waldmann, *Enzyme Catalysis in Organic Synthesis*, Wiley-VCH, **2002**
- V. Gotor, I. Alfonso, E. Garcia-Urdiales, *Asymmetric Organic Synthesis with Enzymes*, Wiley-VCH, **2008**
- E. Garcia-Junceda, *Multi-Step Enzyme Catalysis*, Wiley-VCH, **2008**
- D. Enders, K.-E. Jaeger, *Asymmetric Synthesis with Chemical and Biological Methods*, Wiley-VCH, **2007**
- A.S. Bommarius, B. R. Riebel, *Biocatalysis*, Wiley-VCH, **2004**
- G. Carrea, S. Riva, *Organic Synthesis with Enzymes in Non-Aqueous Media*, Wiley-VCH, **2008**

## Desymmetrizations

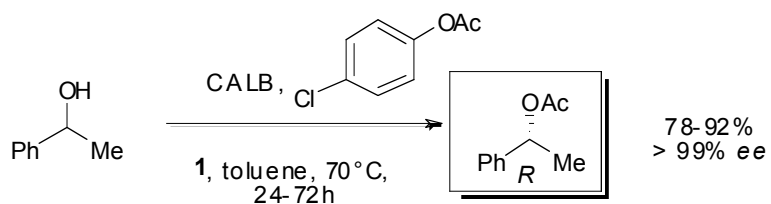
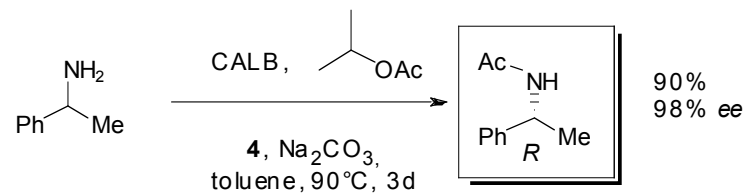
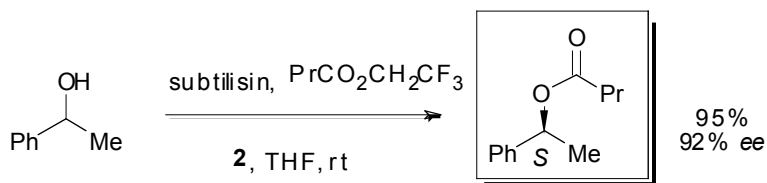
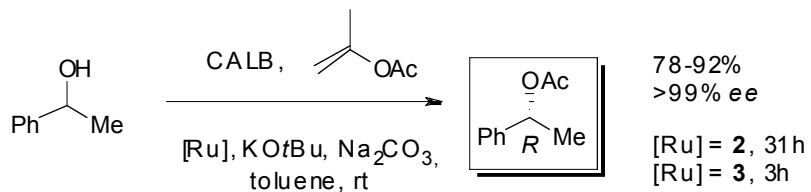


## Dynamic kinetic resolutions

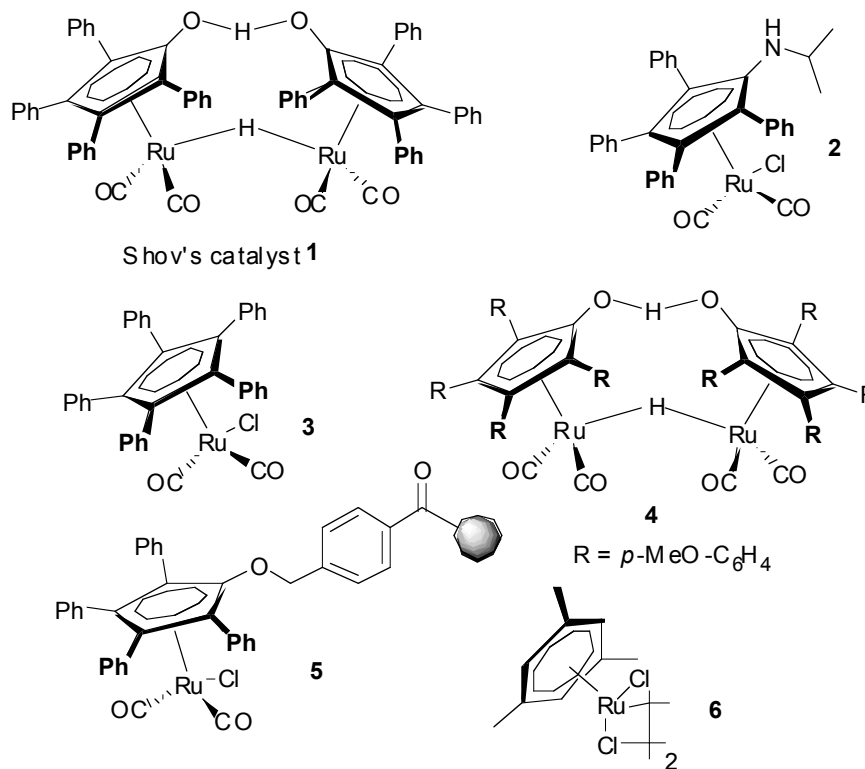
## enzyme-metal combination



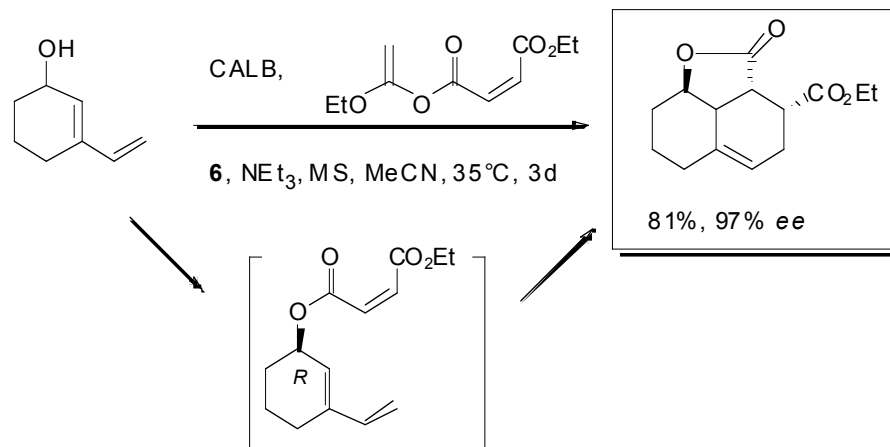
## Ruthenium-catalyzed reactions

used for the production of  $R$ -phenylethanol by DSM

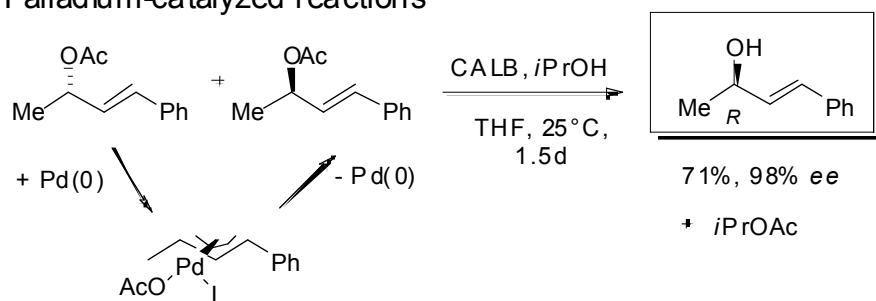
## Ru-catalysts:



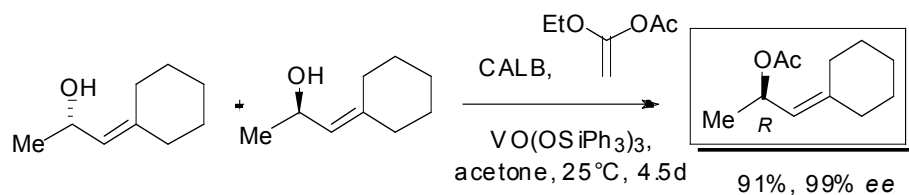
## Tandem-DRK-Diels-Alder reaction



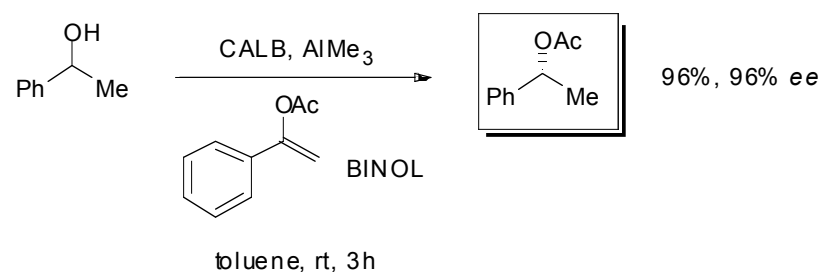
## Palladium-catalyzed reactions



## Vanadium-catalyzed reactions

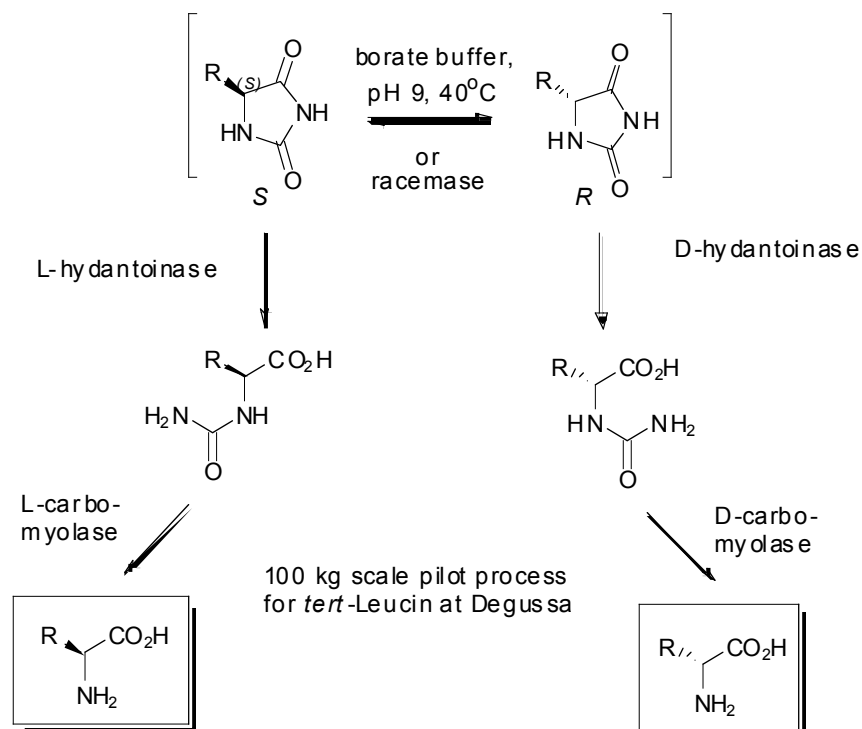


## Meerwein-Ponndorf-Verley-Oppenauer reaction

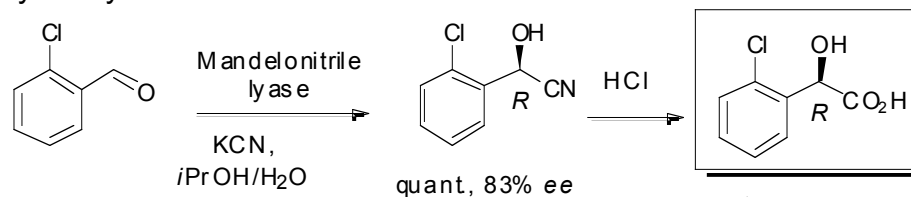


## DRK with enzyme-base combination

## Hydantoinase-carbamylase system



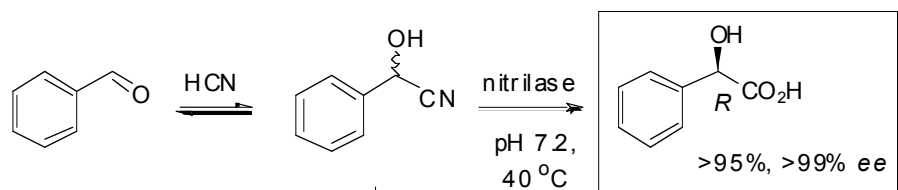
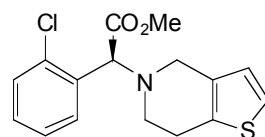
## Cyanohydrine-mediated DRK



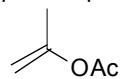
produced by DSM Chemie Linz,  
Nippon Shokubai, Clariant



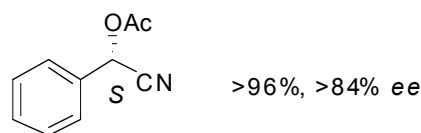
antiplatelet  
clopidogrel (Plavix)



*Pseudomonas  
cepacia* lipase



applied by Lonza, BASF,  
and Mitsubishi Rayon on  
a multiton scale

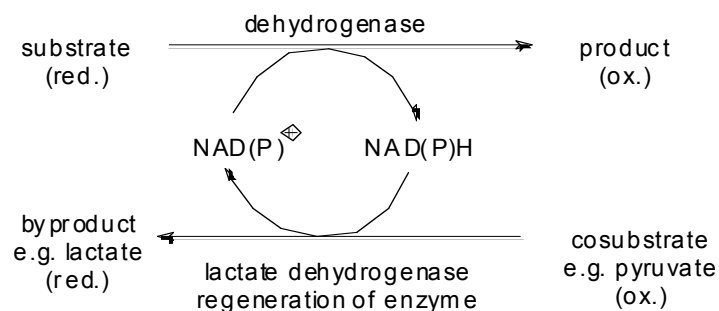


nonselective nitrile hydratase: *Rhodococcus rhodochrous* J1  
- acrylamide production (Nitto process, > 20 000 t/a)  
- nicotinamide synthesis (Lonza, 3000 t/a)

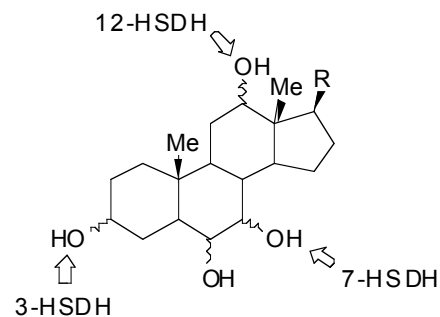
## Oxidations

drawback: co-factor dependence of oxidases/reductases

solutions: - closed-loop systems with an additional enzyme  
for co-factor regeneration  
- electrochemical co-factor recycling  
- application of metals for regeneration  
- living whole cells

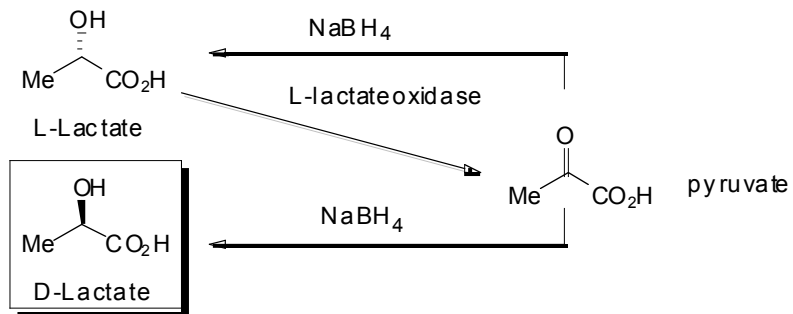


## Oxidations of alcohols and amines

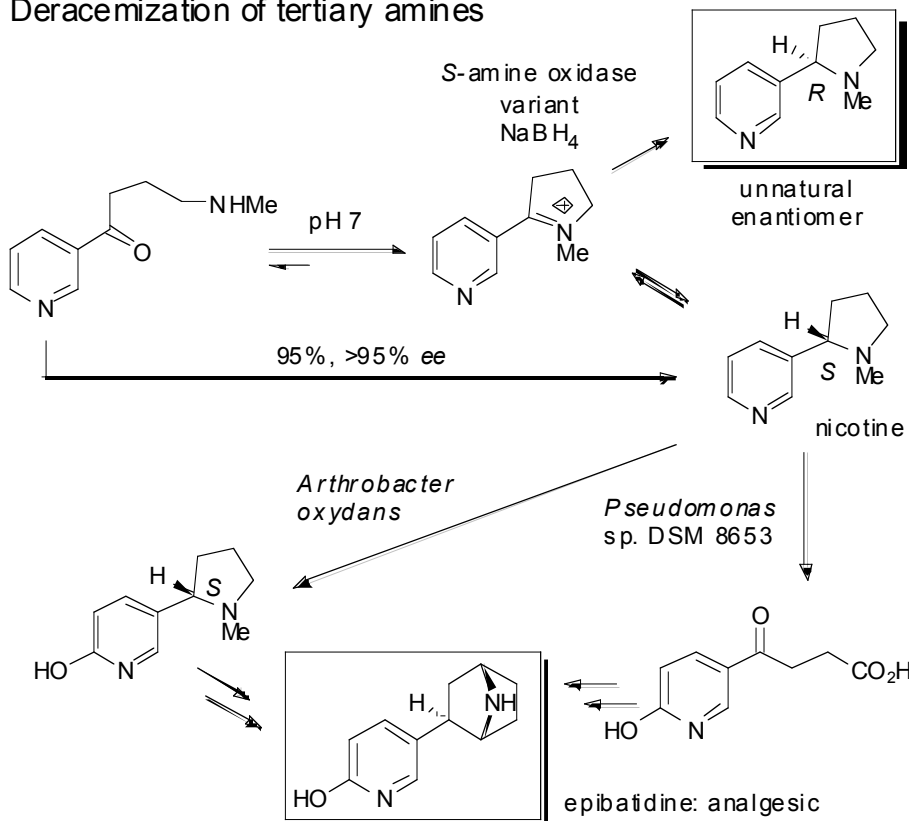


regioselective oxidations of  
bile acid depending on  
hydroxysteroid  
dehydrogenase used

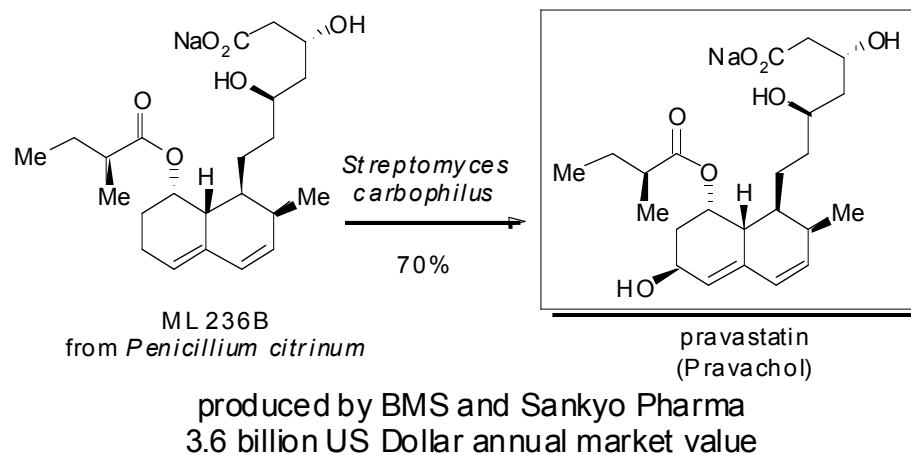
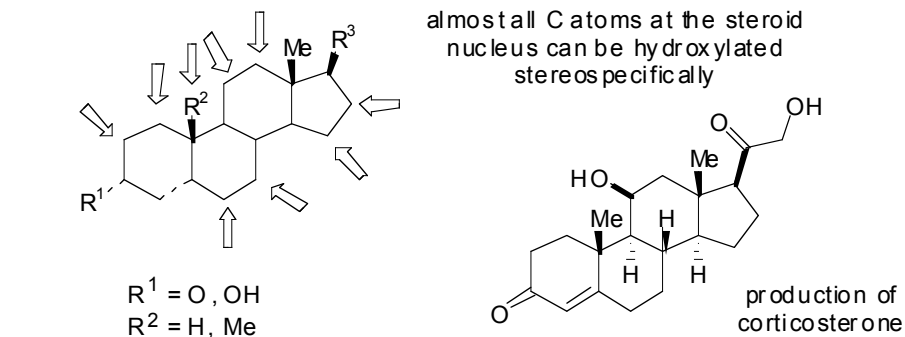
## Deracemization of secondary alcohols



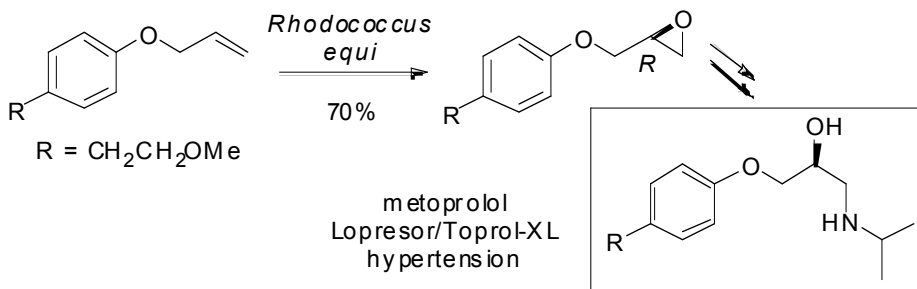
## Deracemization of tertiary amines



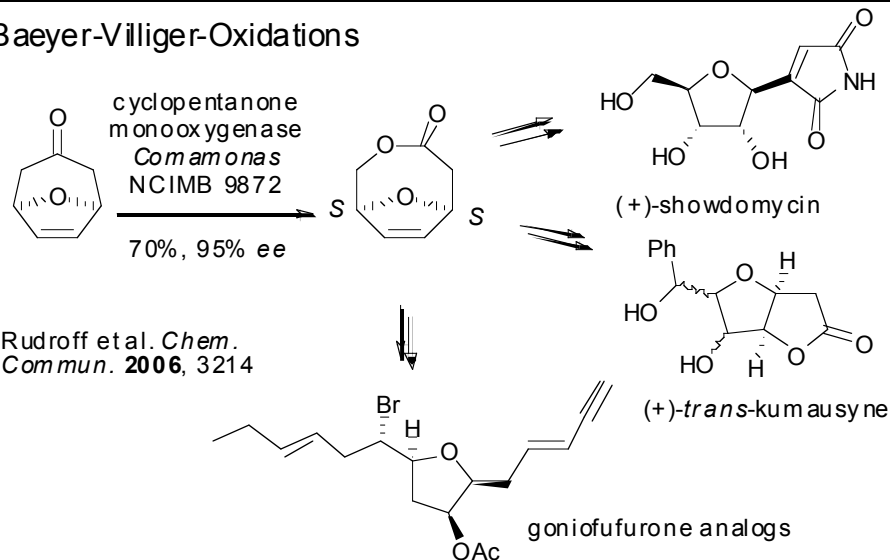
## Oxygenation of nonactivated carbon centers



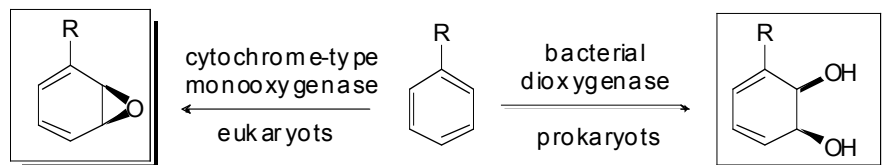
## Epoxidation



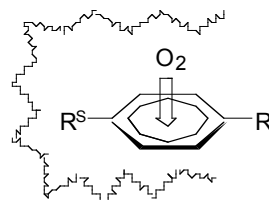
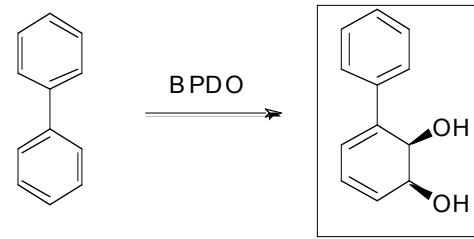
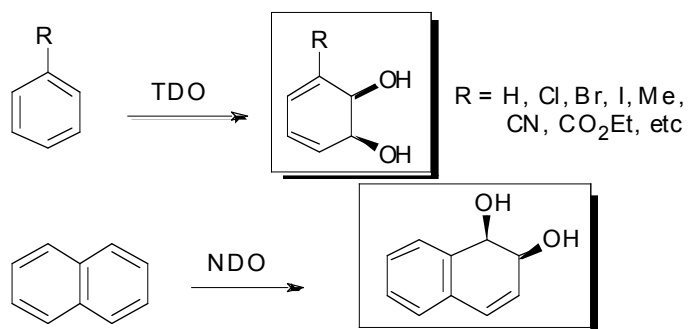
## Baeyer-Villiger-Oxidations

Rudroff et al. *Chem. Commun.* **2006**, 3214

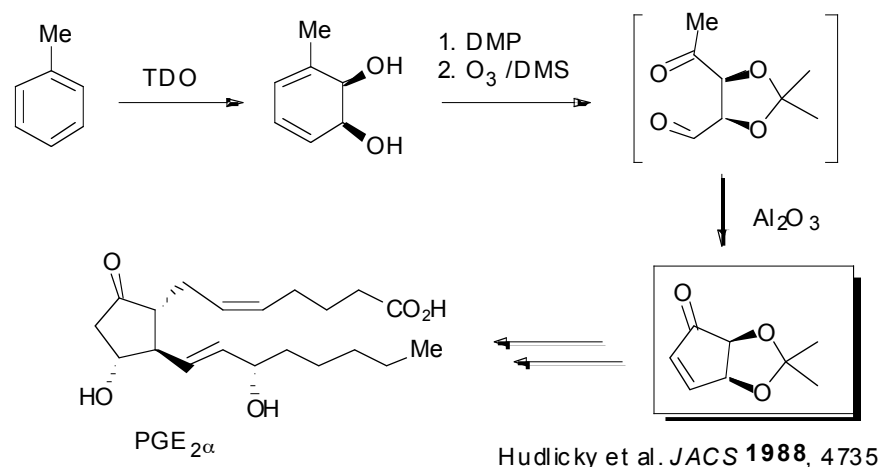
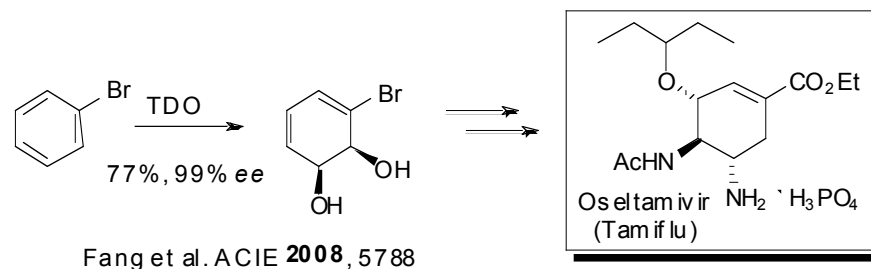
## Aryl dihydroxylations



*ortho* and *meta* hydroxylation occurs using toluene (TDO, *Pseudomonas putida* F39/D), naphthalene (NDO, *P. putida* 119), or biphenyl dioxygenases (BPDO, *Sphingomonas yanoikuyae* B8/36)

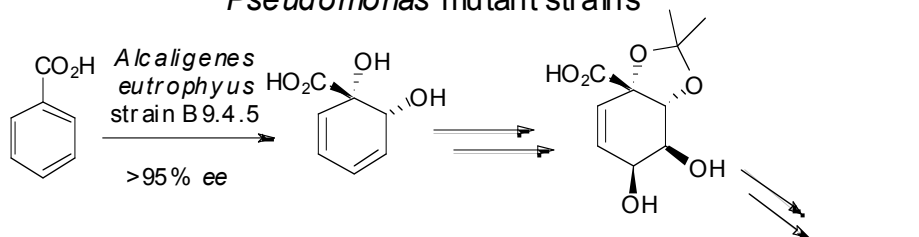


model for predicting the regio- and stereochemical course for the *cis* selective dihydroxylation reaction



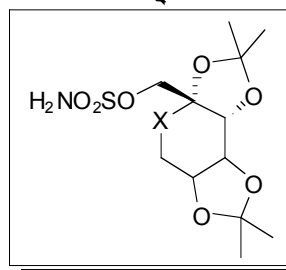


*ipso* and *ortho* dioxygenations possible with *Ralstonia* and *Pseudomonas* mutant strains



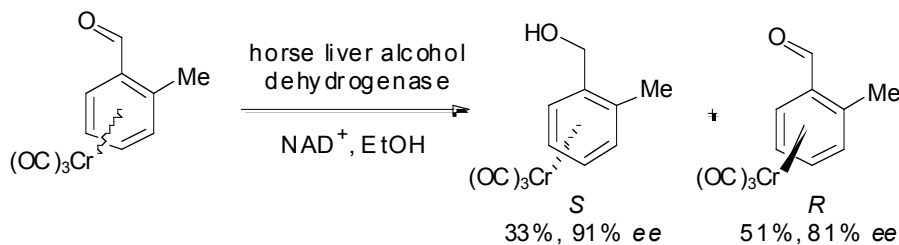
Parker et al. *Synlett* **2004**, 2095

X = O: topiramate  
anti-epilepsy, anti-migrane

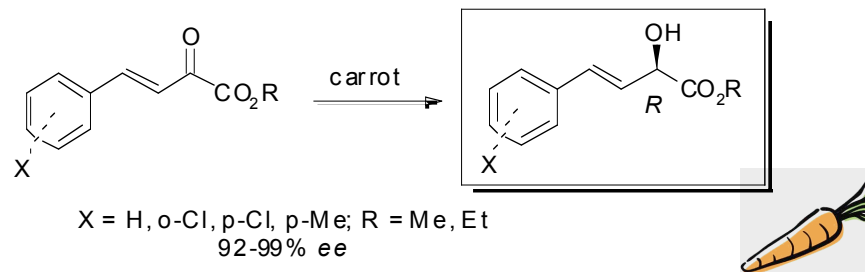
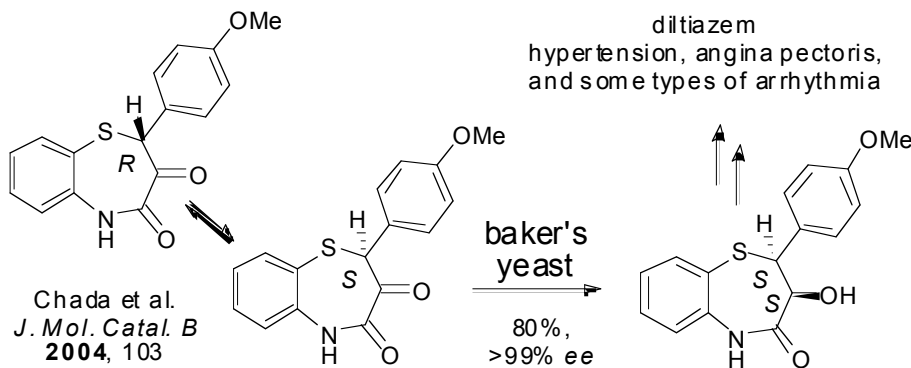


## Reductions

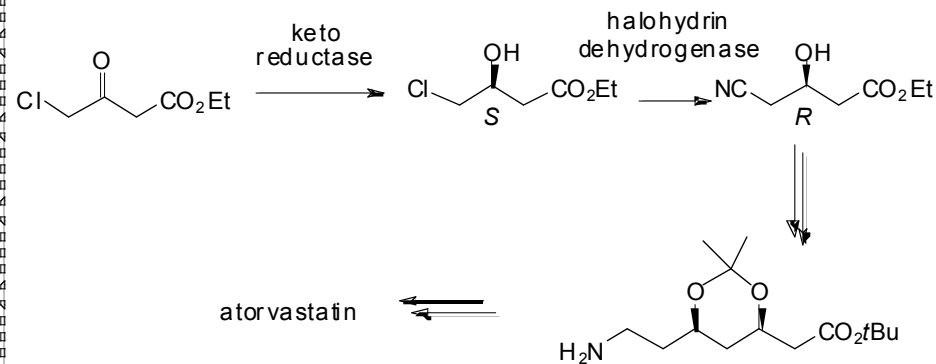
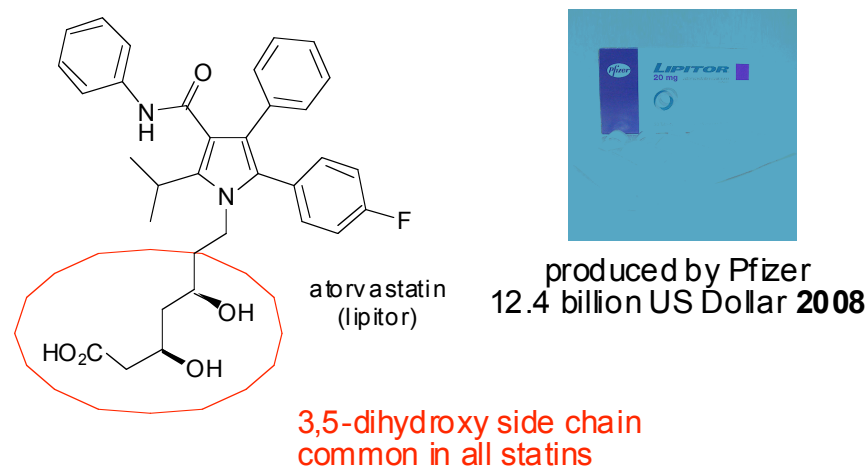
Reduction of aldehydes



Reduction of ketones



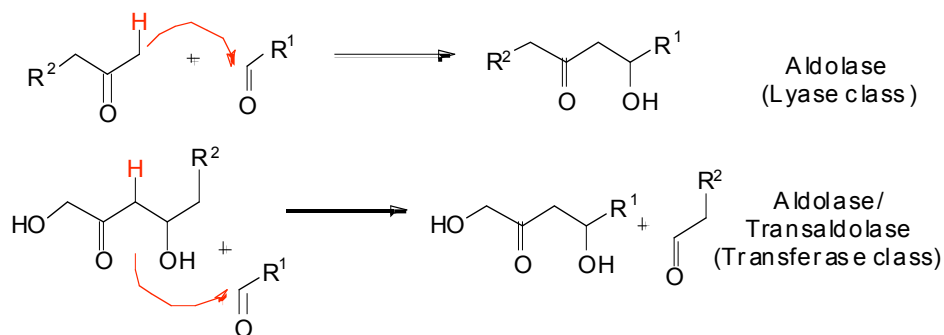
Formation of the 3,5-dihydroxy side chain in statins



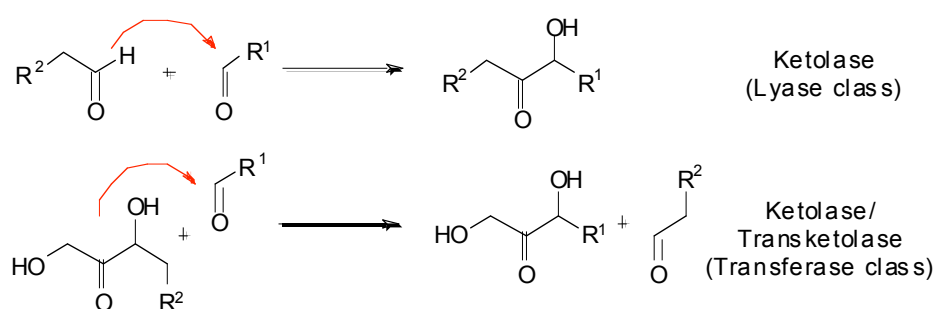
## Carbon-Carbon coupling reactions

stereodivergent product generation possible using  
stereocomplementary enzymes

## Aldol reactions



## Thiamine diphosphate dependent conversions



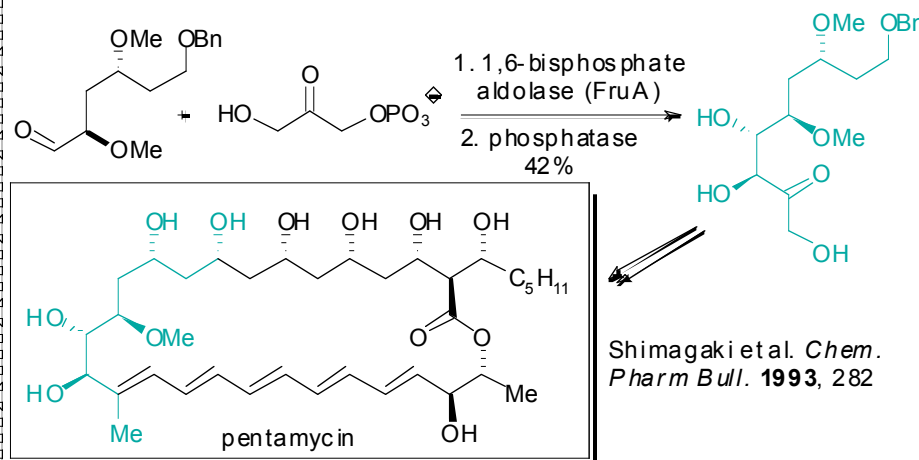
Enzyme classification dependent on the nucleophile:

1. pyruvate-dependent aldolase
2. dihydroxyacetone phosphate (DHAP)-dependend aldolase
3. acetaldehyde-dependent aldolase
4. glycine-dependent aldolase

dihydroxyacetone phosphate (DHAP)-dependend aldolase

-&gt; generation of 2 stereocenters

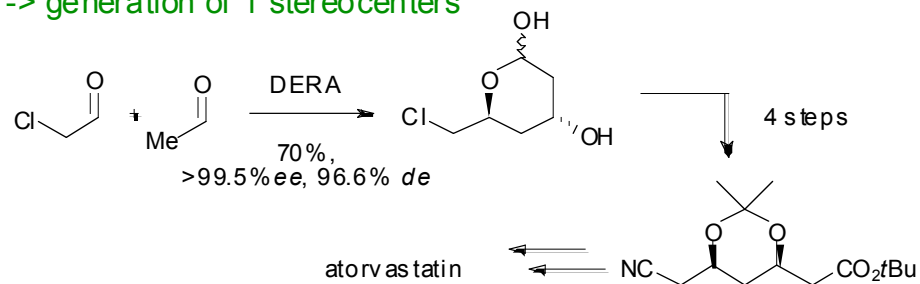
1,6-bisphosphate aldolase (FruA)

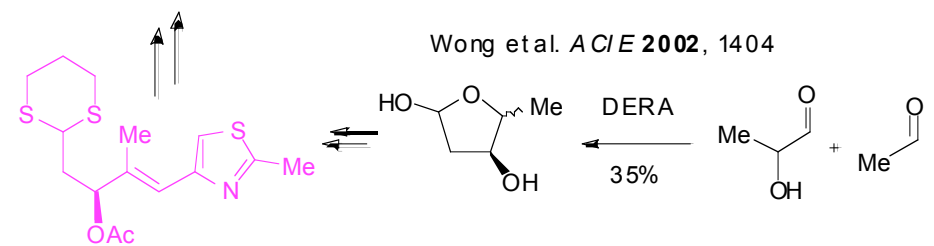
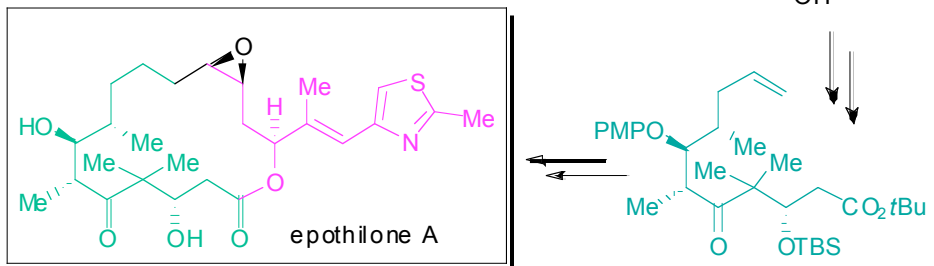
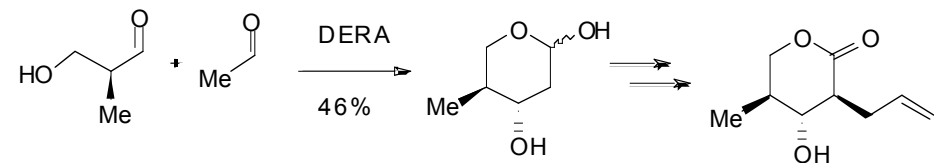


acetaldehyde-dependent aldolase

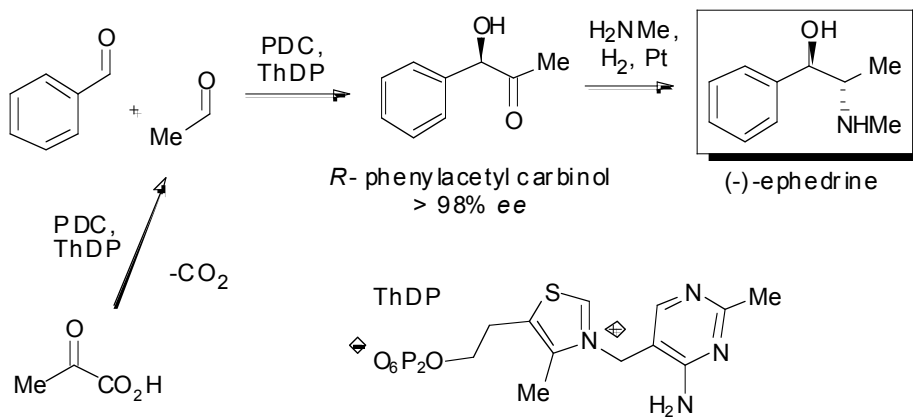
2-deoxyribose-5-phosphate aldolase (DERA)

-&gt; generation of 1 stereocenters





Pyruvate decarboxylase (PDC, thiamin diphosphate depended)



## Total Synthesis of Natural Products

*in vitro* reconstitution of complete biosynthetic pathways

